



Available online freely at www.isisn.org

Bioscience Research

Print ISSN: 1811-9506 Online ISSN: 2218-3973

Journal by Innovative Scientific Information & Services Network



RESEARCH ARTICLE

BIOSCIENCE RESEARCH, 2017 14(3): 626-632.

OPEN ACCESS

Molecular identification of resistant genotype of *Swietenia mahagony* in Egypt using cpDNA trnL-F transgenic spacer region

Eglal, M. Said, Hanafy, Manal M. and Hesham A. El-Aryeb

Horticultural Research Institute, Agricultural Research Center, Giza, Egypt.

*Correspondence: eglal2017@hotmail.com Accepted: 19 Sep. 2017 Published online: 30 Sep. 2017

The genotype of *Swietenia mahogany* plant, grown at Nobarria, Horticulture Research Station, Egypt, showed good tolerance to unfavorable environmental conditions (salinity and calcareous soil) was observed. In this paper, physiological, biochemical and molecular studies were evaluated in order to understand adaptability and tolerance mechanisms of this new genotype. The chloroplast trnL-F region was amplified from extracted total genomic DNA using polymerase chain reaction (PCR) and sequenced. The results showed that the investigated regions of chloroplast genome are variable in the two tested genotypes. These regions served as useful molecular markers in phylogenetic studies or for population studies of *Swatinea mahogany* species.

Keywords: *Swietenia mahogany*, trnL-F, cpDNA, genotype

INTRODUCTION

Genetic diversity is the basis for adaptability and is essential for long term stability of populations and tree breeding for production, whether in plantations or by natural regeneration. It provides the potential for species to resist pests and diseases, and adapt to different environments. Although very strong environmental variation may produce adaptive differences over short distances, despite continued high levels of gene flow (Broadhurst et al. 2008).

Phenotypes of individual plants are determined by genotypes that underlie quantitative traits, environmental conditions, and the interactions between genotype and environment (Wang et al. 2010). The ability of plants to sense environmental changes and produce plastic responses is determined by a portion of the genetic variation, and that plastic phenotypic responses can both provide a buffer against rapid environmental changes and assist rapid adaptation (Nicotra et al. 2010). For any

given plant species, different genotypes may respond similarly or differently to the same environmental changes, or show no response at all, resulting in the differential responses in phenotype (Aspinwall et al. 2015).

Evaluation of chloroplast DNA (cpDNA) diversity in plants is important for characterization of cytoplasm and also for population genetics/phylogeographic analyses. Studies on cpDNA diversity are also important for population genetics and phylogeographic analyses of rare, endemic, and endangered species (Baldwin et al. 1995). The trnL-F intergenic spacer of cpDNA is non-coding characters, and this region is more variable than the coding regions. Some studies on non-coding region of cpDNA showed higher variations and more often mutation than that of coding regions. Therefore, by amplification and direct sequencing of these non-coding regions, the resolution of cpDNA can be increased both for evolutionary studies, and for identifying intraspecific genetic markers (Saiki et al. 1988).

The resolution of cpDNA can be increased both for evolutionary studies, and for identifying intraspecific genetic markers.

Swietenia mahogany belongs to family Meliaceae. Small-leaved mahogany was introduced in India from the West Indies in 1795, and since then has been planted throughout the tropics on a small scale in timber plantations and as an ornamental, occasionally also in tropical Africa. Across their wide geographic distributions, mahogany species appear to be adapted to a wide range of different environments and soils (Navarro et al. 2003).

A pattern resistant genotype of *Swietenia mahogany*, showed good tolerance to unfavorable environmental conditions (salinity and calcareous soil) was observed grown at Noharia, Horticulture Research Station, Egypt, planted almost 15 years ago. In this paper, physiological, biochemical and molecular studies using cpDNA trnL-F region were evaluated in order to understand adaptability and tolerance mechanisms of this resistant genotype.

MATERIALS AND METHODS

This study was carried out at Biotechnology Research Laboratory, Horticulture Research Institute, ARC, Egypt during the period of 2016-2017, on two different genotypes of *Swietenia mahagoni*. One genotype grown at the orchard of Horticultures Institute Research, Giza governorate, and the second genotype grown at Nubaria, Horticulture Research Station, North Tahreer region, Beheira province. Selected trees were about 15 years old.

Soil characteristics

Representative soil samples were collected from each of the studied orchards at depth of 30 to 60 cm and analyzed for Physico-chemical characteristics according to wilde *et al.* (1979) as shown in Table (3).

Photosynthetic Pigments

Photosynthetic pigments (chlorophyll a, b and carotenoids) were measured in *Swietenia mahagoni* leaf by the formula of Lichtenthaler (1987).

Estimation of proline content

Free proline content in the plant tissues was determined following the method of (Bates et al. 1973).

Determination of antioxidant properties

The antioxidant activity of plant methanol extracts was determined based on the radical scavenging ability in reacting with a stable DPPH (2,2-diphenyl-1-picrylhydrazyl) free radical according to Blois (2002).

DNA Isolation

Total DNA was isolated from leaves tissue of the two genotypes as described by Dellaporta *et al.* (1983).

DNA amplification

The chloroplast trnL-F regions were amplified from extracted total genomic DNA using the polymerase chain reaction (PCR) method. The intron of the chloroplast trnL (UAA) gene and the trnL-trnF regions were amplified with primer combinations of (C+D), and (E+F), respectively as recommended by Taberlet et al. 1991 (Table 1 and Fig 1).

DNA Sequencing and Phylogenetic Analysis

PCR products were subjected to gel electrophoresis and cleaned up using a PCR clean-up kit (Promega, USA). Purified PCR products were directly sequenced from one directions, using ABI 3730xl automated DNA sequencer (Applied Biosystems) at GATC Company, USA. The obtained sequence was compared to the sequences in GenBank using the BLAST algorithm to search for close evolutionary relatives.

Gen Bank accession numbers

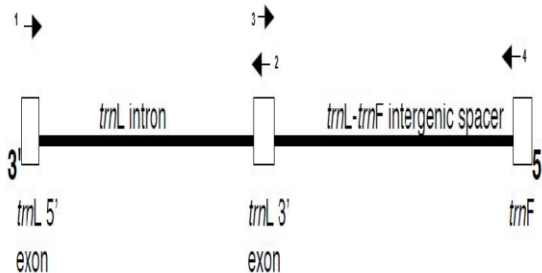
The representative sequence of the new genotype of *Swietenia mahagoni* was deposited in GenBank of National Centre for Biotechnology Information.

Statistical analysis

Statistical analysis was carried out according to Snedecor and Cochran (1976) using analysis of variance and significant difference was determined using L.S.D. values at $P=0.05$, and the distance matrix was represented in a phonogram using UPGMA clustering method (Sneath and Sokal, 1973).

Table 1: Sequences of the trnL-F primers used in this study.

Primer	Sequence 5'-3'
C	CGAAATCGGTAGACGCTACG
D	GGGGATAGAGGGACTTGAAC
E	GGTTCAAGTCCCTCTATCCC
F	ATTTGAACTGGTGACACGAG

**Figure 1: Approximate location of trnL-F primers used in this study**

RESULTS AND DISCUSSION

Photosynthetic Pigments

Chlorophyll (Chl. *a* and *b*), total chlorophyll, carotenoids and total pigments content of leaves were differences between the two investigated genotypes of *Swietenia mahagony*, table (2). Carotenoids and total pigments decreased significantly in tolerance genotype 2, whereas total chlorophyll content revealed insignificant differences between the two genotypes. The decrease in chlorophyll content can be attributed to the sensitivity of this pigment to increasing environmental stresses, especially to drought and salinity (Guerfel et al. 2009). Sharma and Hall (1991), highlighted that saline stress induces degradation of β -carotene, which causes a decrease in the content of carotenoids that are integrated constituents of thylakoid membranes and act in absorption and light transfer to chlorophyll; besides, they protect chlorophyll from photo oxidation, thus degradation in carotenoid synthesis may imply degradation of chlorophylls (Lima et al. 2004).

Proline

The proline content tended to increase in tolerant genotype 2 compared to genotype 1, table (2), although the results have not differed significantly ($p < 0.05$), that proline accumulation was very small in *Swietenia mahagony* species, which indicates that *Swietenia mahagony* has limited capability to synthesize proline as a compatible compound that could increase tolerance or mitigate salt stress effects on this species (Angélica et al. 2011).

Antioxidant effect (DPPH scavenging effect) %

DPPH (2, 2-diphenyl-1-picrylhydrazyl) free radical scavenging method is an easy and rapid method to evaluate free radical scavenging abilities of various samples (Errabii et al. 2006). To prevent and respond to oxidative stress, an anti-oxidative defense system is expressed in the chloroplast, consisting of proteins and scavenging molecules (Froehlich et al. 2003). Data presented in table 2, shows the scavenging effects of extracts on DPPH[•] radicals, the highest value of (92.73%) in genotype 2 was observed, comparison to genotype 1 which reached to (83.44%). Generally with the advent of any stress condition, reactive oxygen species (ROS) are produced. These ROS cause oxidative damage to multiple cellular components like proteins, DNA, RNA and lipids Jacob et al. (1996). All types of abiotic stresses (salinity, chilling, freezing and drought stress) induce oxidative stress in plant cells (De Klerk and Pumisitapon, 2008).

Soil analysis

Data in Table 3 shows that soil texture of Nubaria station is mainly calcareous sandy loam. The values of the EC are 11.7ds/m which is classified very high saline soil while, CaCO₃ values are 30.7, and the pH values are 7.99. The high contents of calcium carbonates causes fixation of phosphorus, low availability of certain micronutrients (B, Fe, Zn, Ni, and Cu), and weak top soil structure Anter et al. (1973).

Table 2: Change in photosynthetic pigments (mg g/FW), antioxidant % and proline content (mg g/FW), in leaves of two different genotypes of *Swietenia mahagony* trees.

Genotype	Chlorophyll a	Chlorophyll b	Total Chlorophyll	Carotenoids	Total pigments	Antioxidant %	Proline
1	0.6269 ^a	0.3085 ^a	0.9354 ^a	6.7884 ^a	7.7238 ^a	83.4489 ^b	0.1546 ^a
2	0.2809 ^b	0.4646 ^a	0.7455 ^a	3.3672 ^b	4.1137 ^b	92.7337 ^a	0.2427 ^a

Means having the same letters in a column were not significantly different at $p < 0.05$

Table 3: Physico-chemical characteristics of the investigated orchard loamy sand soil

Samples	PH	EC ds/m	SP	Soluble anions (mg/l)				Soluble cations (mg/l)				CaCO ₃ %
				CO ₃	HCO ₃	Cl	SO ₄	K ⁺	Na ⁺	Mg ⁺⁺	Ca ⁺⁺	
1	7.96	7.8	42	-	1.2	1.2	4.3	0.5	43.5	11.6	19.4	3.1
2	7.99	11.7	55	-	1.5	105	3.5	0.13	56.3	13.7	24.7	30.7
Particle size distribution (%)												
	Sand			Silt				Clay				
1	85.5			9.5				5				
2	72.5			18.6				8.9				

1 and 2 = soil samples of orchard of Horticultures Research Institute, Giza governorate and Nubaria Horticulture Research Station, Beheira province respectively. EC = Electrical conductivity, PH = Acidity algorithm, ds/m = desicisemen/meters, SP= saturation percentage

Amplification of DNA coding for trnL- F region

The genomic DNA of two genotype of *Swietenia mahagony* was subjected for isolation of the DNA coding for trnL- F (Fig 1). Due to a sequencing problem in the total trnL- trnF region, two primers combinations were used separately. A (C+D) primers amplified the intron of the trnL, while (E+F) primers amplified the partial trnL gene and trnL-trnF regions (Fig 1). PCR amplification of the trnL- trnF region was successfully obtained for (Genotype 2) only, meanwhile amplification was not successful for the second genotype (Genotype 1). The samples had PCR product of about 600bp. and 433bp. with the (C+D) and (E+F) primer sets, respectively (Fig. 2). The PCR product for both primers; (C+D) and (E+F) were directly sequenced from one direction after gel purification

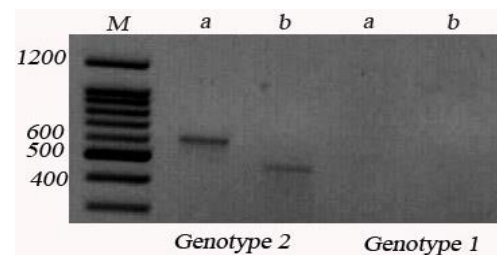


Figure.2. Agarose gel showing amplified trnL intron (a), and trnL-F intergenic spacer (b) for two *Swatinea mahagony* genotypes. Lane M: 100 bp. Ladder

The results showed that the investigated regions of chloroplast genome are variable in the two tested genotypes. These regions served as useful molecular markers in phylogenetic studies or for population studies of *Swatinea mahagony* species

Thus an intraspecific chloroplast DNA polymorphism could result from normal intraspecific variation, which has not been studied for non-coding region, or from intraspecific chloroplast DNA transfer (Taberlet et al. 1991).

The sequences obtained was as follows: Sequence of trnL intron, 600bp.

```

1   GGAACGGAGT TTAGTCATGA TATGGAACCT ACTAAGTGAT AACTTTACAC ATTCAGAGAA
61  ACCCTGGAAT CAAAAATGGG CAATCCTAGA GCCAAATCCT GTTTTACAAG AACAAACAAG
121 GGTTTCAGAAA GCGAAAAAGG GGATAGGTGC AGAGACTCAA TGGAAGCTGT TCTAAGAAAT
181 GGGGTTGACT GCCTTTTTTT TTTTTGGTAA AAAAAGGAAA AAAAATCCTT CTATCAAATA
241 TCAAACTCC ATAAAGGATG AAGGATAAGC GTATATACAC TATGTATACA CAATGAAAAA
301 CTATCTCAA AATGACAACC GAATCCGTAT TTTTTTTTAG GAAAAAAAAA AAAATTGTTA
361 TGAATCAATT CCAAGTTGAA AAAAAAATCA AATATTCATT GATCAAATCA TTCACCCCAA
421 AGTCGGACCA ATCTTTTCTT TTGAAAAACT GATTAACCGG AAAAAATAA AGATAGAGCC
481 CCATTCTACA TGTC AATATCA ATACCGGCAA CAATGAAATT TAAAGTAAAA
    GGAAAAATCCG
541 CCAACTTTAA AAATCGGGAG GGTTC AAGTC CCTCCTATCC CCAACTCTTC GAACTATCCC

```

Sequence of trnL-F region, 433bp.

```

1   GGTCGCGAAT AATCACCATT GTGCTAAATT TCTCCTACCC TCTTTTTGTG TTAGTGGTTC
61  AAAATTCGTT AGGTTTCTCA TTCATCCTAC TCTTTTCCAG ATACAAATGT ATCTGAGCAG
121 AATTTTTTTC TCTTATCACA AGTCGTGTTA TATATATGAT AGACGTACAA ATTAACACCC
181 TTGAGCAAGG AATCCCCAGT TGAATGATGC ACAATTCATA TTATTGCTCA TACTGAAACT
241 TACAAAGTCT TCCTTTTGAA AATTCAAGAA ATGAAATTCC CCGTGCAAGA CTTTAAATAC
301 TTTTTTTTGT CTTTTTAAAT TGACATAGAC CCAAGTCATC TAGTAAAATC AGGATGGTGT
361 GTTGGGAATG GTCGGGATAG CTCAGCTGGT AGAGCAGAGG ACTGAAAATC
    CTCGTGTCAC
421 CAGTAAAAA AAA

```

The chloroplast DNA (cpDNA) *trnT-F* region in land plants consists of the transfer RNA genes trnT UGU, trnL UAA, and trnF GAA arranged in tandem and separated by non-coding spacer regions.

The amplification of the trnL- trnF region in the resistant genotype 2 only, demonstrated that this region may be served as useful molecular markers for stress resistance genotypes.

Sequence analysis and alignment

The sequence of the trnL and trnL-F intergenic spacer of resistant genotype (Genotype 2) was aligned to determine phylogenetic assessment. The trnL region showed 94% identity with *Swietenia mahagoni*, *Khaya grandifoliola* and *Khaya nyasica*, whereas the trnL-trnF region showed 99% identity with *Swietenia mahagoni*, *Khaya nyasica*, *Khaya grandifoliola*, *Dacryodes buettneri*, *Dacryodes buettneri* and *Cedrela odorata*.

As a consequence, the *trnL-F* region, comprising the *trnL* intron and *trnL-F* spacer, has become one of the most widely used accumulation of an increasingly large number of sequences of the *trn (T)-LF* region from a wide range of plants has allowed further study of structures, functions, and evolution in different orders of flowering plants (Bakker *et al.*, 2000). Sequences from the *trnL-F* region (excluding the *trnT-L* region and *trnL* 5' exon) have recently been

used, in combination with those from further chloroplast markers *rbcL* and *matK*, as a source of characters for phylogenetic reconstruction in the tropical flowering plant family Annonaceae Juss. These phylogenies have been used to answer questions about morphological character evolution (Sauquet, 2003), classification, biogeography (Pirie *et al.* 2006). These markers appeared to contain complementary phylogenetic signals, as is expected from different sequences sampled from the plastid genome (Chase and Cox, 1998), and were thus applied in combined analyses.

CONCLUSION

A pattern resistant genotype of *Swietenia mahogany*, showed good tolerance to unfavourable environmental conditions (salinity and calcareous soil) was observed grown at Nobaria, Horticulture Research Station, Egypt, planted almost 15 years ago. In this paper, physiological, biochemical and molecular studies using cpDNA trnL-F region were evaluated in order to understand adaptability and tolerance mechanisms of this resistant genotype .

CONFLICT OF INTEREST

The present study was performed in absence of any conflict of interest.

ACKNOWLEDGEMENT

The author would thank all participants and their parents.

AUTHOR CONTRIBUTIONS

All authors contributed equally in all parts of this study.

Copyrights: © 2017 @ author (s).

This is an open access article distributed under the terms of the [Creative Commons Attribution License \(CC BY 4.0\)](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author(s) and source are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

REFERENCES

- Anter F, Hilal MH, and El-Damaty AH, 1973. A chemical and biological approach towards the definition of calcareous soils. II. Plant growth, P32 and Fe uptake as affected by percentage of carbonate fraction. *Plant Soil*, 39:449-486.
- Angélica, M.C.G., Marina S.S, Maura C., and Cristiane F.T. 2011. Effect of salt stress on nutrient concentration, photosynthetic pigments, proline and foliar morphology of *Salvinia auriculata* Aubl. *Acta Limnologica Brasiliensia*, 23(2): 164-176.
- Aspinwall MJ, Loik ME, Resco De Dios V, Tjoelker MG, Payton PR, and Tissue DT, 2015. Utilizing intraspecific variation in phenotypic plasticity to bolster agricultural and forest productivity under climate change. *Plant, Cell & Environment*, 38, 1752-1764.
- Bakker FT, Culham A, Gomez-Martinez R, Carvalho J, Compton J, Dawtrey R and Gibby M, 2000. Patterns of nucleotide substitution in angiosperm cpDNA trnL (UAA)-trnF (GAA) regions. *Mol Biol Evol*.17(8):1146-55.
- Baldwin BG, Sanderson MJ, Porter JM, Wojcichowski MF, Cambell CS and Donoghue MJ., 1995. The ITS region of nuclear ribosomal DNA: a valuable source of evidence an angiosperm phylogeny. *Annu Miss Bot Gard*, 82:247-277.
- Bates, L.S., R.P. Waldren and I.D. Teare, 1973. Rapid determination of free proline for water stress studies. *Plant Soil*, 39: 205-207.
- Blois MS., 2002. Antioxidant determinations by the use of a stable free radical. *Nature*. 26: 1199-1200.
- Broadhurst, L.M., Lowe, A., Coates, D.J., Cunningham, S.A., McDonald, M., Vesk, P.A. and Yates, C. 2008. Seed supply for broadscale restoration: Maximising evolutionary potential. *Evolutionary Applications* 1: 587–597.
- Chase, M.W. and Cox, AV., 1998. Gene sequences, collaboration and analysis of large data sets. *Australian systematic Botany*: in press.
- De Klerk GJM, and Pumisitapon P., 2008. Protection of in-vitro grown Arabidopsis seedlings against abiotic stresses. *Plant Cell, Tissue and Organ Culture*. 95: 149-154.
- Dellaporta, S. L.; Wood, J. and Hicks, J. B. 1983. A plant DNA mini preparation. Version III. *Plant Mol. Biol.*, Rep.1: 19-21.
- Errabii T, Gandonou CB, Essalmani H, Abrini J, Idaomar M, and Skali-Senhaji N., 2006. Growth, proline and ion accumulation in sugarcane callus cultures under drought-induced osmotic stress and its subsequent relief. *African Journal of Biotechnology*, 5: 1488- 1493.
- Froehlich JE, Wilkerson CG, Ray WK, McAndrew RS, Osteryoung KW, Gage DA, and Phinney BS., 2003. Proteomic study of the Arabidopsis thaliana chloroplastic envelope membrane utilizing alternatives to traditional two-dimensional electrophoresis. *J Proteome Res*. 2:413–425.
- Guerfel M, Baccouri O, Boujnah D, Chaibi W., and Zarrouk M., 2009. Impacts of water stress on gas exchange, water relations, chlorophyll content and leaf structure in the two main Tunisian olive (*Olea europaea* L.) cultivars *Sci Hortic* 119: 257-263.
- Jacob RA, and Burri BJ., 1996. Oxidative damage and defense. *Am J Clin Nutr*. 63: 985–990.
- Lichtenthaler, H.K., 1987. Chlorophylls and carotenoids: pigments of photosynthetic biomembranes. *Methods Enzymol.*, 148: 350-82.
- Lima, MGS., Lopes, NF., Bacarin, MA. and Mendes, CR. 2004. Efeito do estresse salino sobre a concentração de pigmentos e prolina em folhas de arroz. *Bragantia*, 63 (3): 335-340.

- Navarro, C., Wilson, J., Gillies, A. and Hernández, M. 2003. A new Mesoamerican Collection of Big-Leaf Mahogany. In: A.E. Lugo, J.C. Figueroa & M. Alayón (eds.). Big Leaf Mahogany: Genetics, Ecology and Management. Ecological Studies 159, Springer-Verlag New York.. 103–117.
- Nicotra AB, Atkin OK, Bonser SP, Davidson AM, Finnegan EJ, Mathesius U, Poot P, Purugganan MD, Richards CL, Valladares F, and Van Kleunen M, 2010. Plant phenotypic plasticity in a changing climate. Trends in Plant Science, 15: 684-692.
- Pirie, MD, Chatrou, LW, Mols, JB, Erkens, RHJ and Oosterhof, J, 2006. Historical biogeography of two cosmopolitan families of flowering plants. Journal of biogeography. 33, 31-46.
- Saiki RK, Gelfand DH, Stoffel S, Scharf SJ , Higuchi R, Horn GT, MuUis KB, and Erlich HA., 1988. Primer-directed enzymatic amplification of DNA with a thermostable DNA polymerase. Science, 239:487--491.
- Sauquet H., 2003. Androecium diversity and evolution in Myristicaceae (Magnoliales), with a description of a new Malagasy genus, *Doyleanthus* gen. n. J Bot.90(9):1293-305.
- Snedecor, G.M. and Cochran, W.C. 1976. Statistical Methods. 6th Ed. The Iowa State Univ. press. Ames., Iowa, U.S.A.
- Sharma, PK. and Hall, DO. 1991. Interaction of salt stress and photoinhibition on photosynthesis in barley and sorghum. Journal of Plant Physiology, 138(5): 614-619.
- Sneath PHA. and Sokal RR. 1973. Numerical taxonomy: the principles and practice of numerical classification. Freeman WH. San Francisco, California, USA.
- Taberlet P, Gielly L, Pautou G and Bouvet J., 1991. Universal primers for amplification of three non-coding regions of chloroplast DNA. Plant Mol Biol. 17(5):1105-9.
- Wang TL, O'Neill GA, and Aitken SN., 2010. Integrating environmental and genetic effects to predict responses of tree populations to climate. Ecological Applications, 20, 153-163.
- Wilde, S. A., R. B. Corey, J. G. Lyer and G. K. Voigte, 1979. Soil and Plant Analysis for tree culture. Oxford and IBH publishing co., New Delhi, 96-106.