

Available online freely at www.isisn.org

Bioscience Research

Print ISSN: 1811-9506 Online ISSN: 2218-3973 Journal by Innovative Scientific Information & Services Network



RESEARCH ARTICLE BIOSCIENCE RESEARCH, 2019 16(1):287-298.

OPEN ACCESS

Assessment of a sub-chronic consumption of sodium benzoate (E211) on male reproductive functions in Swiss mice

Dalal Redouane, Youcef Bouferkas, Malika Guendouz, Abir Haddi, Nabila Mehedi, Djamel Saidi and Omar Kheroua.

Laboratory of Physiology of Nutrition and Food Safety, Department of Biology, Faculty of Natural and Life Sciences, University Oran 1 Ahmed Ben Bella, Oran, Algeria.

*Correspondence: dalalredouane@gmail.com Accepted: 11 Jan.2019 Published online: 25 Feb. 2019

Sodium benzoate (SB) is a commonly used as a chemical food preservative. Studies on the influence of benzoates on reproductive patterns are very rare in the scientific literature. The present study investigates the effect of sub-chronic consumption of sodium benzoate on male reproductive function in Swiss mice. Thirty Swiss male mice were randomly divided into five groups of 6 mice each. Control group was given drinking water, and treated groups were given sodium benzoate at doses of 0.1, 0.25, 0.5 and 1%. After 13 weeks, we observed a significantly decrease body weight gain, sperm count, sperm motility, normal morphological sperm and serum testosterone levels as well as a significantly increased relative testes weight and impaired testes histology in 0.5 and 1% sodium benzoate treated groups. These findings suggest that excessive consumption of sodium benzoate induces impair spermatogenesis and sperm quality which affects the reproductive performance of male Swiss mice.

Keywords: Sodium benzoate, Testis, Sub-chronic toxicity, Male reproduction, Mice.

INTRODUCTION

Sodium benzoate (E211) is one of the most commonly chemical preservative (Saad et al., 2005; Mpountoukas et al., 2008), used in a variety of foods, beverages and condiments (Zengin et al., 2011) to prevent alteration or degradation caused by microorganisms during storage (Lück, 1985). Sodium benzoate is a salt of benzoic acid, which is used to inhibit the growth of bacteria; it exhibited inhibitory activity against a wide range of fungi, yeasts, molds and bacteria (Mota et al., 2014; Alsudani, 2017).

In addition, sodium benzoate is used to treat hyperammonemia in patients with urea cycle disorders (Leonard et Morris 2002; Scaglia et al., 2004), in order to facilitate an alternative pathway of nitrogen excretion.The joint Expert Committee on Food Additives (JECFA) of the Food and Agriculture Organization of the United Nations (FAO) and the World Health Organization (WHO, 1997) suggested a limit on the amount of sodium benzoate, which is 0 - 5 mg/kg. It is noted that intake estimations from several countries are averaged at 0.18–2.3 mg/kg body weight. However, individuals in China can consume up to 14 mg/kg body weight per day from diet alone (WHO, 1999). The US Food and Drug Agency (FDA) have classified sodium benzoate as "Generally Recognized as Safe" and regulated the concentration of sodium benzoate to 0.1% by weight in food products and 1% concentration in medicines (FAO, 1994; Rothschild, 1990).

However, the consumption of sodium benzoate beyond its acceptable daily intake (ADI) levels may produce toxic consequences in the exposed population. Many studies incriminated sodium benzoate to be responsible for hyperactivity in children (McCann et al., 2007; Beezhold. Johnston and Nochta. 2014), genotoxicity (Loutsidou et al., 2012; Pongsavee 2015), teratogenicity and high mortality in zebrafish embryos (Tsay et al., 2007; Chen et al., In addition, 2009). it can also cause hepatocirrhosis (Kaboglu and Aktac, 2002), muscle damage (Michalak and Qureshi, 1995), cancer (Xing et al., 2000), and leads to the change of morphological structure of lymphocyte and membrane damage in mice (Hu et al., 2008).

Other effects are attributed to sodium benzoate, such as hypersensitivity (Brahmachari and Pahan, 2007; Maier et al., 2010), contact urticaria and contact dermatitis (Andersen, 2006; Sutton and Nixon, 2006), cytostaticity in human lymphocytes (Mpountoukas et al., 2008), inhibition of gluconeogenesis, ureagenesis (Cyr et al., 1991), and fatty acid oxidation (Kalbag and Palekar, 1988).

Nevertheless, so far there are no studies on the effect of sodium benzoate on the testicular toxicity or on the reproductive function. Thus, the purpose of this study is to show the affects of subchronic consumation of sodium benzoate on the reproductive functions in Swiss male mice.

MATERIALS AND METHODS

Chemicals

Sodium benzoate (CAS No. 532-32-1; C7H5O2Na; benzoic acid, sodium salt [E 211 (EU No. Regulation on Labeling of Foodstuffs)]; molecular weight 144.11mol/l) was purchased from Prochima Sigma Tlemcen (Algeria). Triton X-100 and gentian violet were purchased from Sigma (USA), hematoxylin and eosin stains were purchased from Merck (Germany).

Animals and treatments

Male Swiss mice weighting 23.22±0.52g purchased from Pasteur institute, Algiers, Algeria, were kept under proper conditions of ambient temperature and adequate humidity.

Mice were divided into five groups of six animals each. The first group was given drinking water as a control, the second the drinking water containing 0.1% sodium benzoate, the third the drinking water containing 0.25% sodium benzoate, the fourth the drinking water containing 0.5% sodium benzoate, and the fifth the drinking water containing 1% sodium benzoate each for 13 weeks (Chipley, 1983; Toth, 1984). Mice were fed a commercial standard diet (Local Production of Bouzareah, Algiers) and given free access to water all along the experiment. Food and water intake were measured every day and body weight weekly.

The experiments described in this study comply with the current Algerian legislation covering the protection of animals.

Reproductive performance study

After 13 weeks of experiment, groups of sodium benzoate-treated mice, six males per dose group, were mated 1:1 with untreated females for 1 week. Females were then separated and allowed to gestate to term. For females that failed to deliver a litter, this was considered as a sign of male infertility whereas litter delivery indicated male fertility.

Sacrifice of animals and collection of biological samples

After mating, the mice were fasted for 18 hours and killed by cervical dislocation. Testes and epididymis were removed immediately, cleaned of adhering tissues and weighed. Then, they were prepared for fertility evaluation. Right testis for each animal was taken for histopathological examination through light microscopy.

Sperm motility

The left epididymis was excised and placed in a warmed petri dish containing 4 ml of physiological saline solution at 37°C. The tissue was minced with scalpels for approximately 1 min and placed in a 37°C incubator for 15 min prior to determining sperm motility. The suspension was stirred; one drop was placed in a hemocytometer. Under a light microscope (400x magnification), Sperm motility was expressed as percentage of total sperm (Llobet et al., 1995; Yousef and Salama 2009).

Sperm count

The cauda epididymis and the testis were homogenized in 5 or 10 mL of a solution of 0.9% NaCl containing Triton X100 0.5% respectively. The testis and epididymis homogenates were diluted with 1.5 ml of the homogenization solution and spermatozoa were counted at 400x magnification using a Malassez hemocytometer. Five counts *per* sample were averaged (Llobet et al., 1995; Yan et al., 2013).

Sperm morphology

20 µL of suspension were spread onto a glass slide and allowed to dry at room temperature. Once air-dried, the cells were fixed in 96% ethanol for 5 min, stained with 0.5% gentian violet and rinsed with distilled water. 200 spermatozoa were analyzed *per* animal using bright field illumination at final magnification of 1000x (oil immersion). Morphological abnormalities were classified into general categories pertaining to head, mid-piece and tail morphology (Gautam et al., 2010).

Hormone assay

Blood was collected from the retro-orbital venous plexus. Blood serum was separated by centrifugation (3500 rpm for 15 min). Serum testosterone concentrations were measured by a competitive ELISA kit according to manufacturer's instructions (ab108666; Abcam).

Histological analysis

Histological examination of testis was performed. The right testis was fixed in 10% formalin-buffer. Six microns thick paraffin sections were stained with hematoxylin and eosin (H&E) and examined by light microscopy (Optica Axiom 5000, China).

Statistical analysis

Data are expressed as mean values \pm SE. Statistical analysis was performed using statistical test one way ANOVA to find significant difference between values of various parameters recorded for control and treated animals. p<0.05 was considered statistically significant. All statistical analysis were performed using Graph Pad Prism 5 Project software (version 5.01 2007 Graph Pad Software, San Diego, California, USA).

RESULTS

The oral administration of sodium benzoate to the Swiss male mice for 90 days exhibited behavioral and morphological changes like loss of body weight, agitation and aggressiveness.

Body weight and testes weight

Body weight gain was significantly decreased in 1% benzoate group compared with 0.1% group (p < 0.05), as well as in 0.5% (p < 0.01) and 1% (p < 0.001) treated groups compared to control (Table 1).

No significant changes in the absolute testes weight were recorded in all treated groups compared to the control. However, a statistically significant increase of relative testis weight was observed in 1% benzoate group compared with 0.1% group (p < 0.05). The same result was observed in 0.5% (p < 0.05) and 1% (p < 0.01) treated groups compared to the control (Table 1).

Reproductive performance

The result of the fertility test performed on mice Swiss male, treated with sodium benzoate is shown on Table 2. Male mating index was significantly decreased in 0.5% (p < 0.05) and 1% (p < 0.001) treated groups compared to the control values, and in 1% group compared to the 0.1% (p < 0.01). We also noted that females coupled with intoxicated males have a reduced offspring rate compared to females mated with control mice.

Sperm parameters

Table 3 shows the various sperm parameters in the five groups. It reveals that sperm count in testis and epididymis, motility and morphology were decreased in all treated groups compared to control. Significant decrease was observed in the testis and epididymis sperm count of 0.5% (p < 0.01) and 1% (p < 0.001) sodium benzoate treated groups compared to control group. In addition, we observed a significant decrease in sperm epididymis count between the two treated groups 0.1 and 1% (p < 0.01).

Also, a significant decrease in sperm motility was found in 0.5% (p < 0.05) and 1% (p < 0.01) treated groups compared with control, as well as in 1% group compared with 0.1% group (p < 0.05).

In contrary, significant increase was observed in the abnormal sperm rate of 0.5% (p < 0.01) and 1% (p < 0.001) sodium benzoate treated groups compared to the control group, and in 1% group compared with 0,1% group (p < 0.01). Morphological abnormalities involved the sperm head, mid piece and sperm tail (Fig. 1).

Testosterone levels

Serum testosterone level was decreased in all treated groups compared to the control values. However, a significant reduction in serum testosterone level was observed in groups treated with 0.5% and 1% of sodium benzoate compared to control group (p < 0.01) (Fig. 2).

Histological Study

Histological study in the testis of control mice revealed normal structure of interstitial tissue and seminiferous tubules with dynamic spermatogenesis (Fig. 3 A). The germ cells (spermatogonia, primary and secondary spermatocytes, spermatides and spermatozoa) and Sertoli cells within the seminiferous tubules were normal. However, testes of mice treated with sodium benzoate displayed variable degree of histopathological alterations.

We observed dilation in some seminiferous tubules with mild degenerative and necrotic changes, in some of the spermatogonial cells, and detachement germ cells from the irregular basal lamina, in testes of mice of all treated groups (Fig. 3 B, C, D and E). The testes of mice treated with 0.5% of sodium benzoate (Fig. 3 D) showed important dilation in seminiferous tubules, loss germ cells, and decline of spermatogenesis within the individual lumen with absence of spermatozoa in the lumen of seminiferous tubules and beginning dystrophy of the interstitial spaces.

In 1% sodium benzoate treated group, we observed severe degeneration and necrosis of all the seminiferous tubules (Fig. 3 E), disruption in layers germinal epithelium and widening of the interstitial spaces with atrophy of interstitial cells around the seminiferous tubule.

Table 1: Effects of oral ingestion of sodium benzoate on body weight in adult male mice.

Groups concentrations (%)	Control	0.1	0.25	0.5	1
Initial BW (g)	22.22 ± 0.59	22.92 ± 0.84	23.9 ± 0.6	22.18 ± 1.2	24.89±0.42
Final BW (g)	47.16 ± 0.47	43.97±0.61	43.91±0.6	40.55±0.52	41.45±0.71
Body weight gain (g)	24.94±0.55	20.99±1.13	20.00±0.64	18.37±1.33**	16.7±0.71 ^{***, #}
Testes weight (g)	0.233±0.012	0.234±0.51	0.264±0.005	0.256±0.01	0.272±0.01
Testes/BW ratio (%)	0.503±0.02	0.512±0.031	0.593±0.01	0.63±0.019 [*]	0.657±0.02 ^{**, #}

Results are expressed as mean \pm SE (n=6). Significance of the difference was determined using One-Way ANOVA. *p < 0.05, **p < 0.01, ***p < 0.001 significance of difference versus control; # p< 0.05, significance of difference versus 0.1% group.

Table 2. Effect of sodium benzoate on reproductive performance.

Parameters	Control	0.1%	0.25%	0.5%	1%
Mating index § (%)	6/6 (100)	6/6 (100)	6/6 (100)	5/6 (83)*	4/6 (67) ***, ##
Average litter number	9,16±0.30	8.5±0.22	7.5±0.42	5.1±1.08	3.33±1.09
No. of off spring	55	51	45	31	20

§: No. of males producing a pregnant female/No. of males co housed with females;

Results are expressed as mean \pm SE (n=6). Significance of the difference was determined using One-Way ANOVA. *p < 0.05 **p < 0.01, ***p < 0.001 significance of difference versus control; ##p < 0.01, significance of difference versus 0.1% group.

Table 5. Openni parameters in male owiss mile treated with sourdin benzoate.						
Groups concentrations (%)	Control	0.1	0.25	0.5	1	
Count <i>per</i> testis (x10 ⁶)	6.88±0.4	4.33±0.41	3.68±0.31	3.03±.42**	2.43±0.45***	
Count <i>per</i> epididymis (x10 ⁶)	20.7±2.59	14.26±2.34	9.45±0.9	5.91±.53**	3.76±0.52 ^{***, ##}	
Motility (%)	73.91±2.28	64.08±2.44	52.11±2.77	47.47±1.23*	36.57±4.78 ^{***,#}	
Abnormal morphology (%)	9,77±0,76	13,19±0.72	15.17±1.08	23.64±0.95**	30.49±1.27 ^{***, ##}	

Table 3. Sperm parameters in male Swiss mice treated with sodium benzoate.

Results are expressed as mean \pm SE (n=6). Significance of the difference was determined using One-Way ANOVA. *p < 0.05 **p < 0.01, ***p < 0.001 significance of difference versus control; #p < 0.05. ##p < 0.01, significance of difference versus 0.1% group.



Figure 1. Gentian violet staining of sperm from mice treated with Sodium Benzoate under a light microscope with magnification of 1000x.

Abnormal mice sperm morphology classifications; a: normal sperm; b: macrocephalic sperm head; c and d: angulated mid piece; e, f and g: sperm with coiled tail, h: short, i: isolated, j: double.



Figure 2. Change in testosterone in sera of different animal groups.

Results are expressed as mean \pm SE (n=6). Significance of the difference was determined using One-Way ANOVA. **p < 0.01: significance of difference versus control.



Figure 3. Light microscopy of the seminiferous tubules from mice treated with or without SB (A) Section in testis of a control mouse showing normal testicular architecture with an orderly arrangement of germinal cells. (B) Testis of 0.1% BS treated mouse showing detached germ cells from the irregular basal and dilatation of seminiferous tubule (arrow). (C)Testis of 0.25% SB treated mouse showing a degenerated germ cells (arrow). (D and E)Testes of 0.5 and 1% SB treated mice showing degenerated interstitial tissue and degenerated germ cells (arrow).

DISCUSSION

The current study sought to determine the effect of sodium benzoate on reproductive parameters in Swiss male mice.

We observed a decrease in body weight gain, sperm count, sperm motility and serum testosterone levels. However, we showed an increased relative testes weight and abnormal sperm morphology as well as damage in testis structure.

Our study revealed a significant decrease in the body weight gain and significant increase in the relative testes weight in 0.5 and 1% sodium benzoate treated groups. The reduced weight gain may be attributed to reduced food intake (data not shown). This result is consistent with the study of Fujitani et al. (1993) who showed a reduce in average weight of the rats treated with sodium benzoate at a concentration of 2.4%, with increase in relative liver and kidney weight.

In a 90-days study with rats dosed with 0, 1, 2, 4, or 8% sodium benzoate via diet, the mortality in the highest dose group was about 50%, with reduced weight gain, increased relative weights of liver and kidneys along with histopathological changes, of the survivors animals (Deuel et al., 1954). This study conclued that sodium benzoate fed at level of 4 % in the diet have no harmful effect on rats. On the other hand, it has been demonstrated that 4% dose of sodium benzoate in diet results in growth depression within 35 days, in surviving Swiss mice (Toth, 1984).

The loss weight of the gonads, epididymis and accessory sex organs as well as reduced sperm count and epididymal sperm motility, are considered standard criteria for the characterization of toxic agents that may cause fertility problems in the treated subjects (Ban et al., 1995; Queiroz-Neto et al., 1997).

Dietary benzyl acetate, expressed as benzoic acid equivalents (JECFA, 1997), at up to 5% in the diet for 13 weeks, had no effect on the weights of the epididymis, cauda epididymis, or testis, on sperm motility or density, or on the percentage of abnormal sperm in mice of rats (US NTP, 1993). Nevertheless, we observed a significantly reduce in testicular and epididymal sperm counts, with decrease in sperm motility in 0.5 and 1% sodium benzoate treated mice. Reduction in total sperm count reflects that less sperms are produced in testes. These findings are similar to data from Fathabad et al. (2017) who showed that sodium sulfite, another commonly food preservative, affected spermatogenesis, epididymal morphometry, and sperm parameters. There can be several explanations for less sperm production such as loss of spermatogonia, arrest of cell cycle and death of intermediate stages of sperm formation.

In addition, the long-term administration of butyl paraben, at the upper-limit acceptable daily intake (10 mg/kg body weight/day), also caused the reduction of sperm counts in the testis and epididymis (Oishi, 2002a).

Previous work has found parabens to be associated with sperm DNA damage in men as measured by the percentage of DNA in comet tail (Meeker et al., 2010). Methyl and propyl paraben have been shown to affect mitochondrial activity in isolated rat hepatocytes (Nakagawa et Moldeus, 1998), which may be a mechanism of male infertility (Soni et al., 2002; Tavares et al., 2009). Furthermore, the motility of sperm relies on ATP synthesized by oxidative phosphorylation and ceases when the mitochondria become damaged or uncoupled (Jääskaläinen et al., 2003).

Presence of abnormal sperm is another useful indicator of chemical toxicity on the reproductive cells. A high consumption of foods containing preservatives, such as the "Western diet" is associated with poorer sperm count and normal sperm morphology (Liu et al., 2015).

In our present study, a statistically significant increase of sperm abnormalities number was observed in the 0.5 and 1% sodium benzoate treated mice. According to Wyrobek et al., (1983) several kinds of mutations can lead to abnormal sperm morphology. Generally, damage to the sperm cell occurs either by physiological, cytotoxic or genetic mechanism (Otitoloju et al., 2010). This investigation is in accordance with in vitro study conducted by Zengin et al., (2011) who showed that sodium benzoate at very high concentrations (100µg/ml), does have genotoxic effects in human lymphoblastoid cell lines. Recent studies suggest that the sodium benzoate associated with other food additives can contribute to the activation of inflammatory pathways (Raposa et al., 2016), and may have a toxicogenomic effect (Loutsidou et al., 2012).

In a four-generation study with male and female rats, no adverse effects on fertility or lactation were seen after dosing with benzoic acid at up to 1% in the diet (Kieckebusch et Lang, 1960). However, in the present study, crosses between males exposed to sodium benzoate and control male mice with healthy females, showed a significant decrease in mating index and the number of births in 0.5% and 1% treated groups. These results may indicate the inability of a male to fertilized, because of various defects in sperm morphology and/or decrease in the numbers of sperm.

Testosterone is essential for maintaining spermatogenesis and male fertility (Oishi, 2002a; Ahangarpour et al., 2014). Chemicals can damage the structure and function of the epididymis directly or through suppressing Leydia cell testosterone production and can also reduce the number of qualitatively normal sperm entering the epididymis (De Grava et Klinefelter, 2014). In the present study, a significantly decrease of testosterone levels was observed in the 0.5 and 1% sodium benzoate treated mice. Additionally, several studies showed that rodent exposure to butylparaben (Oishi, 2001, 2002a), propylparaben (Oishi, 2002b) and butylated hydroxyanisole (Jeong et al., 2005) induced a decrease in testosterone synthesis and adversely affected the male reproductive function.

Indeed, Jana et al., (2006) suggest that decreased serum testosterone is due to inhibition of testicular steroidogenic enzymes, responsible for the synthesis of testosterone.

In the present study, the dose-levels of sodium benzoate produced adverse effect on testes. Studies on evaluating effects of chemicals on male fertility indicated that testis histopathology is most sensitive parameter for detecting any effect (Mangelsdorf et al., 2003; Dent, 2007).

Histological results revealed damage of the seminiferous tubules together with atrophy of interstitial cells (composed of cells Leydig) and inhibition of spermatogenesis in the testis of mice orally exposed to sodium benzoate to 0.25, 0.5 and 1% doses. Testicular damages are characterized by dilatation of some siminiferious tubules in all treated groups and loss of germ cells in 0.5% as well as extensive disruption in 1% treated group. This finding is in agreement with the study that indicates that sodium metabisulfite at 260 mg/kg/day induced many (SMB) histological and biochemical alterations in the testicular tissue of rats (Fathabad et al., 2017).

Leydig cells have a direct role in the regulation of spermatogenesis by producing hormones, specifically testosterone. Reduction in testosterone causes regression of the epididymal epithelium and reduces the androgen-dependent facets of sperm maturation (De Grava and Klinefelter, 2014). O'donnell et al., (1996) suggest that testosterone affects the adhesives function between round spermatids and Sertoli cells, leading to the sloughing of round spermatids for epithelium. It has been confirmed that in the absence of testosterone, progressive germ cell degeneration beguins during stage VII of the spermatogenic cycle (Sharpe, 1994).

Our findings clearly indicate that sodium benzoate has toxic effects on the testis physiology and morphology of Swiss Albino mice when administered at the high dose of 1% equivalent to 1017.31 mg.kg⁻¹.day⁻¹ and middle dose of 0.5% corresponding to 522.6 mg.kg⁻¹.day⁻¹. These doses level were in excess of the ADI of sodium benzoate (0-5 mg kg-1 body weight). In fact, no one knows exactly how much benzoate is ingested through dietary regimen, and sodium benzoate was the first chemical preservative permitted in food for human consumption in the U.S. in 1908 and continues to be used in a large number of foods (Jay, 1992). Furthermore, the international program on chemical safety assured no adverse effects in humans at doses of 647-825 mg/kg body weight per day (Nair, 2001). In China, the maximum permitted utilization of benzoates in different types of food ranges from 0.2 to 2.0 g/kg [GB2760-2014].

Because the level of chemicals in food and also the food consumption pattern are different from country to country, it is necessary in developing countries to conduct their own exposure assessment using domestic data, and compare the local exposure data with the safe intake level developed by international expert groups (Chen, 2004).

CONCLUSION

In conclusion, the present data demonstrate that sub-chronic ingestion of sodium benzoate, in drinking water is able to impair male reproductive health in male mice, at 0.5 and 1%. Above the ADI, sodium benzoate possesses adverse effects. It is very important for reproductive health to carry out surveys among the population to estimate their daily intake of additives, since processed and packaged foods have become a convenient choice for today's busy families.

CONFLICT OF INTEREST

The authors declared that present study was performed in absence of any conflict of interest.

ACKNOWLEGEMENT

This research was supported by the Directorate General for Scientific Research and Technological Development (DGRSDT, MESRS, Algeria).

AUTHOR CONTRIBUTIONS

DR performed the experiments, data analysis and also wrote the manuscript. YB, MG, AH performed animal treatments and tissue collection, NM provided scientific advice, DS and OK contributed to the experimental design and to the manuscript revision. All authors read and approved the final version.

Copyrights: © 2019 @ author (s).

This is an open access article distributed under the terms of the **Creative Commons Attribution License (CC BY 4.0)**, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author(s) and source are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

REFERENCES

- Ahangarpour A, Oroojan AA, Radan M, 2014. Effect of aqueous and hydro-alcoholic extracts of lettuce (Lac- tuca sativa) seed on testosterone level and spermatogenesis in NMRI mice, Iran J Reprod Med. 12:65–72.
- Alsudani A, 2017. In vitro antifungal effect of potassium sorbate and sodium benzoate on the growth of fungi causing sinusitis. Afr J Microbiol Res.11:232-236.
- Andersen A, 2006. Final report on the safety assessment of benzaldehyde. Int. J. Toxicol. 25:11–27.
- Ban Y, Komatu KM, Inagaki S, Nakatsuka MH, 1995. Testicular spermatid and epididymal sperm head count as an indicator for reproductive toxicity in rats. Exp. Anim. 44: 315-322.
- Beezhold BL, Johnston CS, Nochta KA, 2014. Sodium benzoate rich beverage consumption is associated with increased reporting of ADHD symptoms in college students: a pilot investigation. J Atten Disord. 18:236-241.
- Brahmachari S, Pahan K, 2007. Sodium benzoate, a food additive and a metabolite of cinnamon, modifies T cells at multiple steps and inhibits adoptive transfer of experimental allergic encephalomyelitis, J. Immunol. 179:275–283.
- Chen J, 2004. Challenges to developing countries after joining WTO: Risk assessment of chemicals in food. Toxicology. 198:3-7.
- Chen Q, Huang NN, Huang JT, Chen S, Fan J, Li

C, Xie FK, 2009. Sodium benzoate exposure downregulates the expression of tyrosine hydroxylase and dopamine transporter in dopaminergic neurons in developing zebrafish. Birth Defects Res B Dev Reprod Toxicol.89:85-91.

- Chipley JR. Sodium benzoate and benzoic acid, 1983. In: A.L. Branen, P.M. Davidson (eds.), Antimicrobials in foods. M. Decker, New York, pp. 11–35.
- Cyr DM, Egan SG, Brini CM, Tremblay GC, 1991. On the mechanism of inhibition of gluconeogenesis and ureagenesis by sodium benzoate. Biochem Pharmacol.42:645–654.
- De Grava Kempinas W, Klinefelter GR, 2014. Interpreting histopathology in the epididymis. Spermatogenesis. 4:1-12.
- Dent MP, 2007. Strengths and limitations of using repeat-dose toxicity studies to predict effects on fertility. Regul Toxicol Pharmacol. 48:241-258.
- Deuel HJ Jr, Alfin-Slater R, Weil CS, Smyth HF, 1954. Sorbic acid as a fungistatic agent for foods. 1. Harmlessness of sorbic acid as a dietary component. Food Res.19:1–12.
- Fathabad AA, Shekarforoush S, Hoseini M, Ebrahimi Z, 2017. Attenuation of Sulfite Induced Testicular Injury in Rats by Zingiber officinale Roscoe. J Diet Suppl. 18:1-12.
- Food and Agriculture Organization of the United Nations World Health Organization (FAO/WHO), 1994. Summary of evaluations performed by the Joint FAO/WHO Expert Committee on Food Addivites (JECFA). International Life Sciences Institute, Washington.
- Fujitani T, 1993. Short-term effect of sodium benzoate in F344 rats and B6C3F1 mice. Toxicol Lett. 69:171–179.
- Gautam D, Sharma G, Goyal RP, 2010. Evaluation of Toxic Impact of Tartrazine on Male Swiss Albino Mice. Pharmacology online. 1:133-140.
- Hu M, Wang J, Cai J, Wu Y, Wang X, 2008. Analysis of sodium benzoate biotoxicity by atomic force microscope, Shengwu Gongcheng Xuebao, Chin J Biotechnol. 24:1428–1432.
- Jana K. Jana S, Samanta PK, 2006. Effects of Chronic Exposure to Sodium Arsenite on Hypothalamic Pituitary- Testicular Activities in Adult Rats: Possible Anestrogenic Mode of Action. Reprod Biol Endocrinol. 4:9.
- Jay JM, 2005. Modern Food Microbiology. Van Nostrand Reinhold, New York, 1992.

- Jeong SH, Kim BY, Kang HG, Ku HO, Cho JH, 2005. Effects of butylated hydroxyanisole on the development and functions of reproductive system in rats. Toxicology. 208:49–62.
- Joint FAO/WHO Expert Committee on Food Additives (JECFA), 1962. Evaluation of the Toxicity of a Number of Antimicrobials and Antioxidants: Sixth Report of the Joint FAO/WHO Expert Committee on Food Additives. World Health Organization (WHO), Geneva, Switzerland, WHO Technical Report Series No. 228. p104.
- Joint FAO/WHO Expert Committee on Food Additives (JECFA), 1997. Evaluation of Certain Food Additives and Contaminants, Forty-sixth Report of the Joint FAO/ WHO Expert Committee on Food Additives, World Health Organization (WHO), Geneva, Switzerland. WHO Technical Report Series 868.
- Jääskaläinen EL, Teplova V, Andersson MA, Andersson LC, Tammela P, Andersson MC, Pirhonen TI, Saris NE, Vuorela P, Salkinoja-Salonen MS, 2003. In vitro assay for human toxicity of cereulide, the emetic mitochondrial toxin produced by food poisoning Bacillus cereus. Toxicol In Vitro.17:737–44.
- Kaboglu A, Aktac T, 2002. A study of the effects of sodium benzoate on the mouse liver, Biologia. 57:375–382.
- Kalbag SS, Palekar AG, 1988. Sodium benzoate inhibits fatty acid oxidation in rat liver: effect on ammonia levels. Biochem Med Metab Biol. 40:133–142.
- Kieckebusch W, Lang K, 1960. Die Verträglichkeit der Benzoesäure im chronischen Fütterungsversuch, Arzneimittel-Forschung.10:1001–1003.
- Leonard JV, Morris AA, 2002. Urea cycle disorders. Semin Neonatol.7:27–35.
- Liu CY, Chou YC, Chao JC, Hsu CY, Cha TL, Tsao CW, 2015. The association between dietary patterns and semen quality in a general Asian population of 7282 males. PLoS One.10:1-12.
- Llobet JM, Colomina MT, Sirvent JJ, Domingo JL, Corbella J, 1995. Reproductive toxicology of aluminum in male mice. Fundam Appl Toxicol.25:45-51.
- Loutsidou AC, Hatzi VI, Chasapis CT, Terzoudi GI, Spiliopoulou CA, Stefanidou ME, 2012. DNA content alterations in Tetrahymena pyriformis macronucleus after exposure to food preservatives sodium nitrate and

sodium benzoate. Acta Biol Hung.63:483-489.

- Luck E, 1985. Chemical preservation of food. Zentralbl Bakteriol Mikrobiol Hyg B.180:311– 318.
- Maier E, Kurz K, Jenny M, Schennach H, Ueberall F, Fuchs D, 2010. Food preservatives sodium benzoate and propionic acid and colorant curcumin suppress Th1-type immune response in vitro. Food Chem Toxicol.48:1950–1956.
- Mangelsdorf I, Buschmann J, Orthen B, 2003. Some aspects relating to evaluation of the effects of chemicals on male fertility. Regul Toxicol Pharmacol.37:356-369.
- McCann D, Barrett A, Cooper A, Crumpler D, Dalen L, Grimshaw K, Kitchin E, Lok K, Porteous L, Prince E, Sonuga-Barke E, Warner JO, Stevenson J, 2007. Food additives and hyperactive behaviour in 3year-old and 8/9-year-old children in the community: a randomised, double-blinded, placebo-controlled trial, Lancet.370:1560– 1567.
- Meeker JD, Ehrlich S, Toth TL, Wright DL, Calafat AM, Trisini AT, 2010. Semen quality and sperm DNA damage in relation to urinary bisphenol A among men from an infertility clinic. Reprod Toxicol.30:532–539.
- Michalak A, Qureshi A.I, 1995. Free and esterified coenzyme A in the liver and muscles of chronically hyperammonemic mice treated with sodium benzoate. Biochem Mol Med.54:96–104.
- Mota FJ, Ferreira IM, Cunha SC, Beatriz M, Oliveira PP, 2003. Optimisation of extraction procedures for analysis of benzoic and sorbic acids in foodstuffs. Food Chem.82:469–473.
- Mpountoukas P, Vantarakis A, Sivridis E, Lialiaris T, 2008. Cytogenetic study in cultured human lymphocytes treated with three commonly used preservatives. Food Chem Toxicol.46:2390–2393.
- Nair B, 2001. Final report on the safety assessment of benzyl alcohol, benzoic acid, and sodium benzoate. Int J Toxicol.20:23–50.
- Nakagawa Y, Moldeus P, 1998. Mechanism of phydroxybenzoate ester-induced mitochondrial dysfunction and cytotoxicity in isolated rat hepatocytes. Biochem Pharmacol.55:1907–1914.
- O'Donnell L, McLachlan RI, Wreford NG, De Kretser .M, Robertson DM, 1996. Testosterone withdrawal promotes stage-

specific detachment of round spermatids from the rat seminiferous epithelium. Biol Reprod. 55:895–901.

- Oishi S, 2002 (a). Effects of butyl paraben on the male reproductive system in mice. Arch Toxicol.76:423–429.
- Oishi S, 2001. Effects of butylparaben on the male reproductive system in rats. Toxicol Ind Health.17:31–39.
- Oishi S, 2002 (b). Effects of propyl paraben on the male reproductive system. Food Chem Toxicol.40:1807–13.
- Otitoloju AA, Obe IA, Adewale OA, Otubanjo OA, Osunkalu VO, 2010. Preliminary Study on the Induction of sperm head abnormalities in mice, mus musculus, exposed to radiofrequency radiations from global system for mobile communication base stations. Bull Environ Contam Toxicol.84:51–54.
- Pongsavee M, 2015. Effect of sodium benzoate preservative on micronucleus induction, chromosome break, and Ala40Thr superoxide dismutase gene mutation in lymphocytes. BioMed Res Int .2015:5.
- Queiroz-Neto A, Mataqueriro MI, Santana AE, Alessi AC, 1997. Toxic effects of Annona squamosa seed extract in rats and swine. Rev Bras Toxicol.10:11- 15.
- Raposa B, Pónusz R, Gerencsér G, Budán F, Gyöngyi Z, Tibold A, Hegyi D, Kiss I, Koller Á, Varjas T, 2016. Food additives: Sodium benzoate, potassium sorbate, azorubine, and tartrazine modify the expression of NFκB, GADD45α, and MAPK8 genes. Physiol Int. 103:334-343.
- Rothschild DL Jr, 1990. The Food Chemical News Guide guide to the cur- rent status of food additives and color additives.Washington, DC: Food and Chemical News.
- Rotstein, Saad B, Bari MF, Saleh MI, Ahmad K, Talib MK, 2005. Simultaneous determination of preservatives (benzoic acid, sorbic acid, methylparaben and propylparaben) in foodstuffs using high-performance liquid chromatography. J Chromatogr A. 1073:393– 397.
- Safety evaluation of certain food additives, 1999. WHO Food Additives Series, Geneva, No. 42.
- Saad B, Bari MF, Saleh MI, Ahmad K, Talib MK, 2005. Simultaneous determination of preservatives (benzoic acid, sorbic acid, methylparaben and propylparaben) in foodstuffs using high-performance liquid chromatography. J Chromatogr A. 1073:393–

397.

- Scaglia F, Carter S, O'Brien WE, Lee B, 2004. Effect of alternative pathway therapy on branched chain amino acid metabolism in urea cycle disorder patients. Mol Genet Metab.81:79– 85.
- Sharpe RM, 1994. Regulation of spermatogenesis. In E. Knobil, J.D. Neill (eds.), The physiology of reproduction. Raven Press, New York. pp. 1363–1434.
- Soni MG, Taylor SL, Greenberg NA, Burdock GA, 2002. Evaluation of the health aspects of methyl paraben: a review of the published literature. Food Chem Toxicol. 40:1335–1373.
- Sutton T, Nixon R, 2006. Allergic contact dermatitis to sodium benzoate chloroacetamide in a sorbolene lotion. Australas J Dermatol. 47:209–210.
- Tavares R.S., Martins FC, Oliveira PJ, Ramalho-Santos J, Peixoto FP, 2009. Parabens in male infertility-is there a mitochondrial connection?. Reprod Toxicol. 27:1–7.
- Toth B, 1984. Lack of tumorigenicity of sodium benzoate in mice, Fundam. Appl. Toxicol. 4:494–496.
- Tsay HJ, Wang YH, Chen WL, Huang MY, Chen YH, 2007. Treatment with sodium benzoate leads to malformation of zebrafish larvae. Neurotoxicol Teratol. 29:562–569.
- US NTP, 1993. Toxicology and carcinogenesis studies of benzyl acetate (CAS No. 140-11-4) in F344/N rats and B6C3F1 Mice (feed studies). Technical Report Series No. 431, Research Triangle Park, NC, US Department of Health, Education and Welfare, National Institutes of Health, National Toxicology Program.
- Wyrobek AJ, Gordon LA, Burkhart JG, Francis MW, Kapp RW Jr, Letz G, 1983. An evaluation of the mouse sperm morphology test and other sperm tests in non-human mammals. A report of the US environmental protection agency gene-toxicology program. Mut Res.115:1-72.
- Xing WJ, Que LC, Wen W, 2000. Detecting mutagenic effects of pesticides and food additives by using plant SCE. Acta Acad Med Nei Mongol. 22:141–144.
- Yan L, Bai XĽ, Fang ZF, Che LQ, Xu SY, Wu D, 2013. Effect of different dietary omega-3/omega-6 fatty acid ratios on reproduction in male rats. Lipids Health Dis. 12:33.
- Yousef MI, Salama AF, 2009. Propolis protection from reproductive toxicity caused by

aluminium chloride in male rats. Food Chem Toxicol. 47:1168-1175.

Zengin N, Yüzbaşıoğlu D, Unal F, Yılmaz S, Aksoy H, 2011. The evaluation of the genotoxicity of two food preservatives: sodium benzoate and potassium benzoate. Food Chem Toxicol. 49:763–769