



Available online freely at www.isisn.org

Bioscience Research

Print ISSN: 1811-9506 Online ISSN: 2218-3973

Journal by Innovative Scientific Information & Services Network



RESEARCH ARTICLE

BIOSCIENCE RESEARCH, 2019 16(2): 1251-1257.

OPEN ACCESS

Inheritance of resistance to downy mildew (*Peronosclerospora maydis*) in crossing of Madura Maize Plant (*Zea mays* L.)

Achmad Amzeri *, Kaswan Badami and Gita Pawana.

Faculty of Agriculture, Trunojoyo Madura University, Bangkalan, East Java 69162, Indonesia

*Correspondence: aamzeri@gmail.com Accepted: 09 Apr. 2019 Published online: 06 May. 2019

Hybridization of Back cross is one method to get varieties that are resistant to downy mildew. The purpose of this study was to obtain information on inheritance characteristics of downy mildew resistance. This research was conducted at the experiment center of Agro-Technology Study Program of Agriculture Faculty, University of Trunojoyo Madura. Research of Assessment of resistance to Downy Mildew used a randomized block design with 18 treatments (P_1 , P_2 , F_1 , F_2 , BC_1P_1 and BC_1P_2 in three sets of crosses, namely LGL x Mdr-3, T12 x Mdr-1 and E02 x Mdr-2) and three replications so there were 54 experimental units. Identification of polymorphic RAPD markers for endurance to downy mildew through Bulk Segregant Analysis (BSA) was done by amplifying the DNA in the resistant pool and susceptible pool. The random primers used were 120 primers from 6 operon groups, namely OPA, OPB, OPC, OPD, OPF and OPG. The results showed that the inheritance pattern of maize genetic resistance to downy mildew followed a segregation pattern of 3:1 with a degree of dominance between -1 and 0, and was controlled by incomplete partially negative dominant gene. OPC-07 was a marker that was linkage close to the resistance to downy mildew with a genetic distance of 1.9 cM.

Keywords: inheritance, downy midew, madura maize, back cross, bulk segragant analysis

INTRODUCTION

Madura Island is an island located in the East Java region of Indonesia which has a crop area for maize of approximately 360,000 hectares (30% of the area of maize in East Java), but productivity at the farm level is still low at an average of 2.7 tons per hectare (BPS, 2015). These results are very low compared to other maize-producing regions. The low productivity of maize on the island of Madura is caused by low soil fertility and not yet assembled superior maize for the island of Madura (Amzeri et al., 2011).

Increased productivity can be done by assembling superior varieties that have high production and resistant to the main diseases of maize plants. Downy mildew is a major disease in maize plants caused by the fungus

Peronesclerospora. Downy mildew caused a decrease in yield of 10% to 90% (Talanca, 2010) even reaching 100% (Soebandi et al., 1996; Yasin et al. 2008).

Assembling varieties that have high production and resistance to downy mildew can be done by crossing maize plants that have high production with plants that are resistant to downy mildew (Amzeri, 2015). The method of crossing that can be used to produce varieties resistant to disease is back cross (Poespodarsono, 1998; Mangendidjojo, 2003; Amzeri, 2015). The back cross method has produced many varieties that are resistant to disease. In addition, molecular markers can be used as a selection aids (MAS = Marker Assisted Selection), where selection is based on plant genetic traits without

environmental influences, so that plant breeding is faster, right, save money and time (Pabendon et al., 2007; Kumar and Gurusubramanian, 2011).

The purpose of this study was to obtain information on inheritance characteristics of downy mildew resistance. Information about inheritance of a character has an important meaning in determining plant breeding strategies so that breeding programs to improve the desired character are more effective.

MATERIALS AND METHODS

Genetic Material

This research was conducted at the experiment center and biotechnology laboratory of Agro-Technology Study Program of Agriculture Faculty, University of Trunojoyo Madura, Indonesia, in July 2017-June 2018. The parents of maize used were parents resistant and not resistant to downy mildew. The genetic material evaluated were P₁, P₂, F₁, F₂, BC₁P₁ and BC₁P₂ in three crossing sets namely LGL x Mdr-3, T12 x Mdr-1 and E02 x Mdr-2.

Field Experiments and Assessment of Resistance to Downy Mildew

This research used a randomized block design with 18 treatments (P₁, P₂, F₁, F₂, BC₁P₁ and BC₁P₂ in three crossing sets namely LGL x Mdr-3, T12 x Mdr-1 and E02 x Mdr-2) and three replications so there were 54 experimental units. Each experimental unit consists of 400 plants planted with a spacing of 20 cm x 70 cm. waxy corn is a plant that is susceptible to downy mildew planted as a spreader plant (spreader row) as many as three rows on the edge and block of plant population tested. Inoculation was done by spraying suspension spores on plants aged 7, 9 and 11 days after planting, both on the spreader plant lines and on the plant population tested. Observation of the occurrence of downy mildew is done when plants are 21, 28 and 35 days after planting. The number of sample plants observed in P₁, P₂ and F₁ (40 sample plants), BC₁P₁ and BC₁P₂ (80 sample plants) and F₂ (200 sample plants). Resistance to downy mildew is tested based on the percentage of attacks : score 0 = very resistant (0-10%), score 1 = resistant (>10-20%), score 2 = rather resistant (>20-40%), score 3 = susceptible (>40-60%), score 4 = very susceptible (60-100%) (Talanca, 2009).

The degree of dominance is measured based on the parents middle value (P₁ and P₂) and F₁

using the formula for estimating the potential ratio (hp) (Griffing, 1950).

$$hp = \frac{(F_1 - MP)}{1/2(HP - LP)}$$

where : hp = value of potential ratio; HP = value of highest parent average; LP = value of lowest parent average; MP = the average value of the two parents.

Based on the value of potential ratios can be predicted the degree of dominance as follows:

- hp = 0 = additive gene action
- 1 = hp = +1 = perfect dominant
- 1 < hp < 0 = incomplete partially negative dominant gene
- 0 < hp < 1 = incomplete partially positive dominant gene
- 1 < hp < 1 = over dominant

The segregation pattern on F₂ was tested using the chi-square test (Suryo, 2008).

$$\chi^2 = \frac{(O_i - E_i)}{E_i}$$

where : O_i = number of phenotypes observation to-i; E_i = number of phenotype to-i based on hypothesis

DNA preparation

The DNA of F₂ plant groups from the three crosses that resistant (score 0) and very susceptible (score 4) to downy mildew each was isolated following the procedure of Daryono and Natsuaki (2002). The DNA of each group was combined proportionally based on the concentration level to form 1 DNA pool.

DNA amplification

DNA amplification followed the method of Te-Chato et al., (2005). The amplification reaction was done using 20µl of the solution that consist of master mix 11 µl, primer 2,2 µl, DNA 3 µl and dH₂O/1XTE buffer 3,8 µl. The tube containing a mixture of PCR was rotated in a centrifuge with 8000 rpm for 30 seconds to 1 minute. Furthermore, amplification was done in the PCR machine for approximately 4 hours 30 minutes with the program as follows:one pre PCR cycle (92°C, 2 minute), 40 cycles {denaturation (92°C, 1 minute), annealing (37°C,1 minute), elongation (72°C, 2 minute) }, post elongation (72°C, 10 minute). The amplification results were visualized by agarose gel electrophoresis.

Linkage Analysis of RAPD Marker with resistance to downy mildew

Identification of polymorphic RAPD markers for resistance to downy mildew was done by

amplifying resistant DNA pools and susceptible DNA pools using random primers. The random primers that were used 120 primers from 6 operon groups, namely OPA, OPB, OPC, OPD, OPF and OPG. Each group consists of 20 primers with 10 base lengths. Primers that capable to amplifying DNA in the resistant pool were applied to resistant and susceptible parents. From the specific polymorphic markers of the resistant parent obtained, then it was applied to the population F_2 for analysis of linkage resistance to downy mildew. The linkage test used 120 plant populations in three crosses. Linkage analysis between RAPD markers and resistance control was done with the MAPMAKER computer application program.

RESULTS AND DISCUSSION

In three crosses, the rate of downy mildew attack in Family F_1 was between the values of the mid-parents and the resistant parents (Table 1). Schematically the relative position of F_1 for both parents is shown in Figure 1. Takdir et al., (2003) conducted a cross between resistant and susceptible maize produced the rates of downy mildew attack in the F_1 Family whose value was between the middle values of parents and resistant parents. The value of the potential ratio in the three crosses has a value between -0.384 to -0.652. The value of the potential ratio (hp) which is in the range of -1 and 0 indicates that the character is controlled by incomplete partially negative dominant gene (Petr and Frey, 1966).

The segregation pattern in F_2 population given an indication that the resistant character shows a simple genetic inheritance pattern and follows the pattern of segregation of Mendel's law (Table 2). The chi-square test (χ^2) for the degree of suitability of the F_2 population segregation ratio for various hypothetical ratios based on two

endurance classes shows that the segregation pattern of the three F_2 populations matches the 3:1 ratio. At the LGL x Mdr-3 crossing had a value of $\chi^2 = 0.0061$ with an opportunity value (P) of 0.95-0.90. T12 x Mdr-1 crossing had a value of $\chi^2 = 0.2960$ with an opportunity value (P) of 0.70-0.50. E02 x Mdr-2 crossing had a value of $\chi^2 = 2.0575$ with an opportunity value (P) of 0.20-0.10.

Population analysis of back cross (BC_1P_1 and BC_1P_2) show that crossing of LGLxMdr-3, T12xMdr-1 dan E02xMdr-2, back cross to male parent (resistant/ BC_1P_1) and back cross to female parent (susceptible/ BC_1P_2) had a value that matches the 1: 1 ratio at the 0.05 level. The population of BC_1P_1 which derived from a crossing between F_1 and P_1 (resistant parent) approaches the resistant parent. In contrast, the population of BC_1P_2 which derived of crossing between F_1 with P_2 (susceptible parent) approaches the susceptible parent. This event was suitability with Mendel Laws of 1 that in the formation of gametes the two genes that are pairs will be segregated into two daughter cells (Stansfield, 1991).

The results of amplification of the resistant DNA pool and susceptible DNA pool using 120 random primers were obtained that 46 primers that could amplify the resistant and susceptible DNA pool. These primers were used to identify 4 F_2 plants in each pool (4 F_2 plants of resistant and 4 F_2 plants of susceptible). The results of this study indicate that OPC-7 ('GTCCCGACGA') resulted in the application of bands present to resistant parents, bulk resistant and 4 resistant F_2 plants. This band was not found in susceptible parents, susceptible parent bulk and 4 susceptible F_2 plants. This marker had around 475 base pairs (bp) (Figures 5 and 6). OPC-07 was a marker that was linkage close to the resistance to downy mildew with a genetic distance of 1.9 cM.

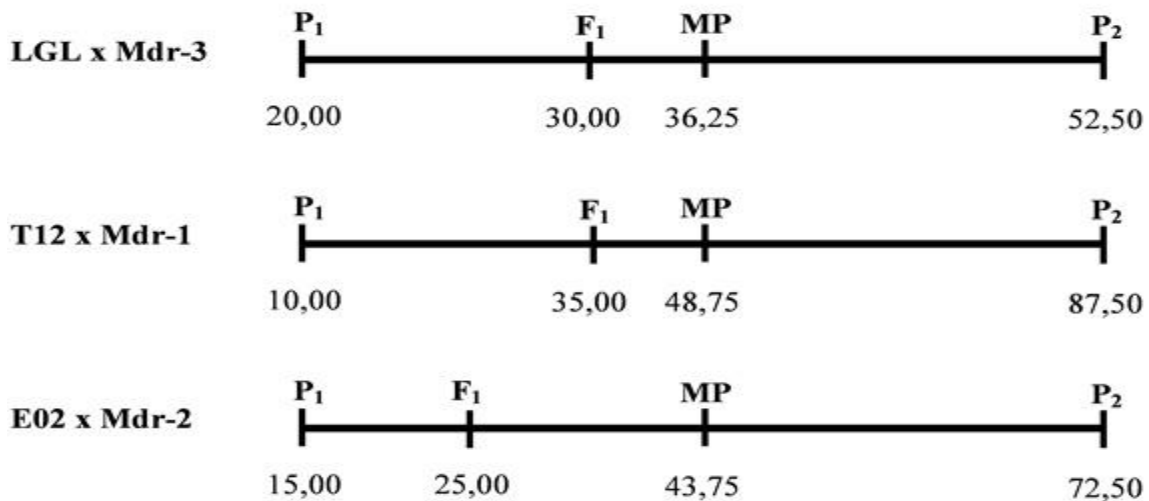
Table 1. The rate downy mildew attack on P_1 , P_2 , F_1 dan potensi rasio (hp)

Family	Attack rate		
	LGL x Mdr-3	T12 x Mdr-1	E02 x Mdr-2
P_1	20,000	10,000	15,000
P_2	52,500	87,500	72,500
F_1	30,000	35,000	25,000
hp	-0,384	-0,355	-0,652

Table 2. Segregation of resistance to downy mildew in crossing of LGL x Mdr-3, T12 x Mdr-1 and E02 x Mdr-2.

Crossing	Genera- tion	Number of plants				Expected ratio	χ^2	(P)
		Observation		Expected				
		Resistant	Susceptible	Resistant	Susceptible			
LGL x Mdr-3	P ₁	32	8	-	-			
	P ₂	19	21	-	-			
	F ₁	28	12	-	-			
	F ₂	143	57	143,50	56,50	3:1	0,0061	0,95-0,90
	BCP ₁	55	25	56,00	24,00	1:1	0,0596	0,80-0,70
	BCP ₂	51	29	51,20	28,80	1:1	0,0729	0,80-0,70
T12 x Mdr-1	P ₁	36	4	-	-			
	P ₂	5	35	-	-			
	F ₁	26	14	-	-			
	F ₂	138	62	141,50	58,50	3:1	0,2960	0,70-0,50
	BCP ₁	51	29	52,00	28,00	1:1	0,0549	0,90-0,80
	BCP ₂	43	37	41,20	38,80	1:1	0,1621	0,70-0,50
E02 x Mdr-2	P ₁	34	6	-	-			
	P ₂	11	29	-	-			
	F ₁	30	10	-	-			
	F ₂	132	68	141,24	58,76	3:1	2,0575	0,20-0,10
	BCP ₁	59	21	60,00	20,00	1:1	0,0667	0,80-0,70
	BCP ₂	44	36	44,5	35,50	1:1	0,0629	0,80-0,70

$$\chi^2_{.05} = 3.841 \text{ and } \chi^2_{.01} = 6,635$$

**Figure 1. Schematic of the relative position of the F₁ middle value of both parents**

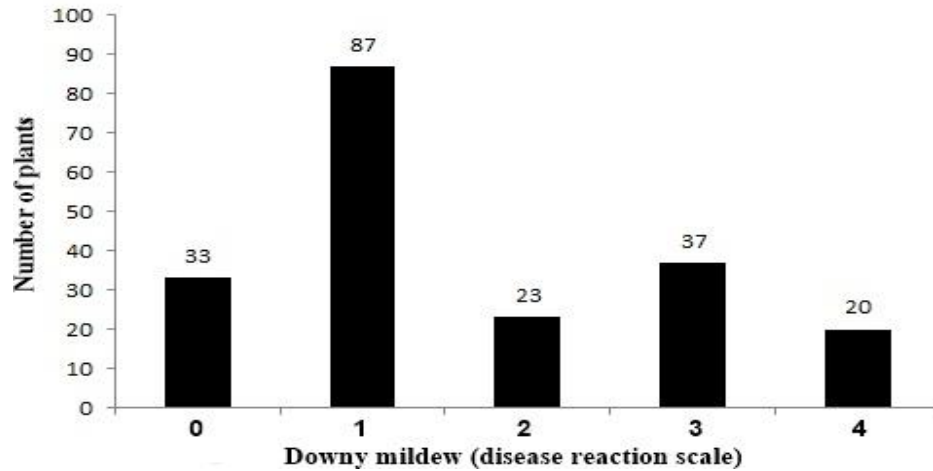


Figure 2. Distribution of F₂ plants as a crossing result of resistant maize plants (LGL) with susceptible maize plants (Mdr-3) based on the level of resistance to mildew.

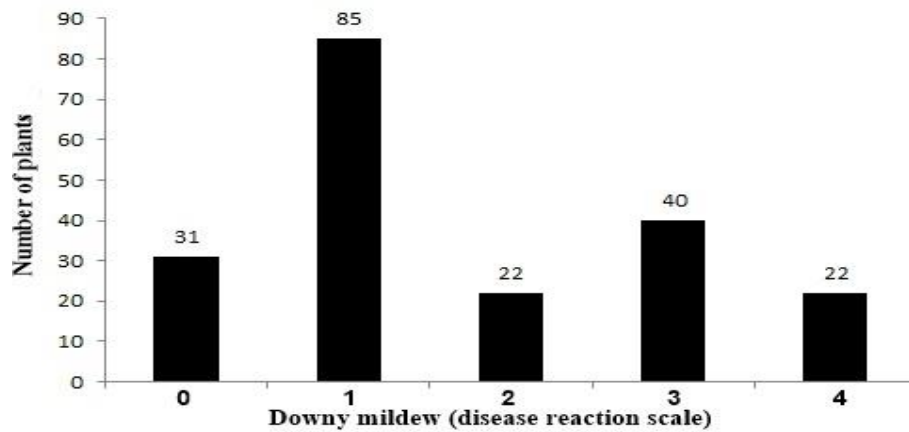


Figure 3. Distribution of F₂ plants as a crossing result of resistant maize plants (E02) with susceptible maize plants (Mdr-2) based on the level of resistance to mildew.

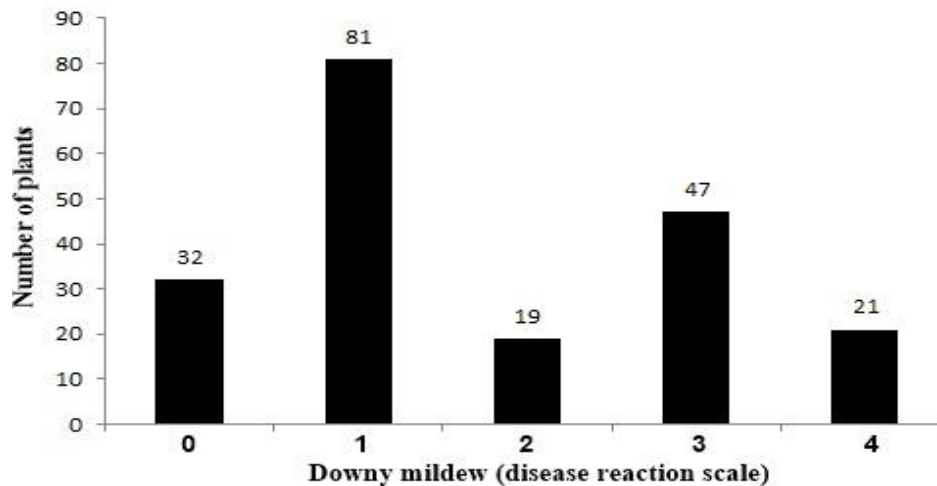


Figure 4. Distribution of F₂ plants as a crossing result of resistant maize plants (T12) with susceptible maize plants (Mdr-1) based on the level of resistance to mildew.

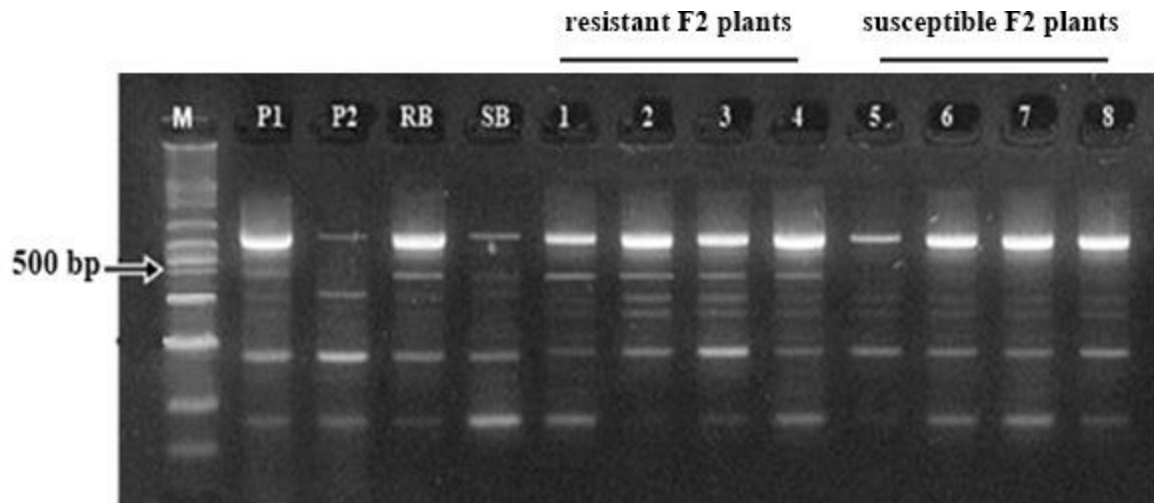


Figure 5. The profile of the DNA fragment amplified with OPC-7 primer in the F₂ population derive from the crossing of LGL x Mdr-3

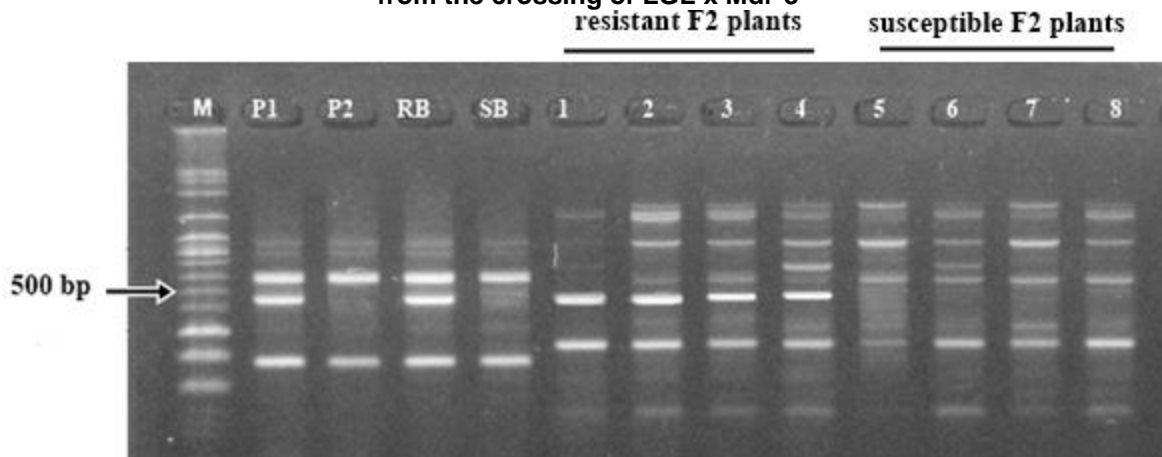


Figure 6. The profile of the DNA fragment amplified with OPC-7 primer in the F₂ population derive from the crossing of T12 x Mdr-1

CONCLUSION

The results showed that the inheritance pattern of maize genetic resistance to downy mildew followed a segregation pattern of 3:1 with a degree of dominance between -1 and 0, and was controlled by incomplete partially negative dominant gene. OPC-07 was a marker that was linkage close to the resistance to downy mildew with a genetic distance of 1.9 cM.

CONFLICT OF INTEREST

The authors declared that present study was performed in absence of any conflict of interest.

ACKNOWLEDGEMENT

This research was supported by a grant from

the Applied Product Research (2017-2018) of Ministry of Research and Technology of Higher Education, Indonesia.

AUTHOR CONTRIBUTIONS

All authors contributed equally in all parts of this study.

Copyrights: © 2019 @ author (s).

This is an open access article distributed under the terms of the [Creative Commons Attribution License \(CC BY 4.0\)](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author(s) and source are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or

reproduction is permitted which does not comply with these terms.

REFERENCES

- Amzeri A, Indradewa D, Daryono BS, and Rachmawati D. 2011. Phenetic and Genetic Relationships among Madura Local Maize (*Zea mays* L.) Revealed by Morphological Characters and RAPD Markers, *Biota*, 16(2), 227-235.
- Amzeri A, 2015. Plant Breeding Basics, UTM Press, Bangkalan Madura. Indonesia. pp : 223.
- BPS, 2015. East Java Maize Productivity, <http://Jatim.bps.go.id/> tgl 23 juli 2016.
- Daryono BS and Natsuaki KT, 2001. Application of random amplified polymorphic DNA markers for detection of resistant cultivars of melon (*Cucumis melo*) against Cucurbitaceae viruses, In II International Symposium on Cucurbits 588 (pp. 321-329).
- Griffing B, 1950. Analysis of quantitative gene action by constant parent regression and related techniques, *Genetics*, 35(3) : 303.
- Kumar NS and Gurusubramanian G, 2011. Random amplified polymorphic DNA (RAPD) markers and its applications, *Sci Vis*, 11(3) : 116-124.
- Mangoendidjojo W, 2003. Plant Breeding Basics, Kanisius, Yogyakarta, Indonesia. pp : 194.
- Pabendon MB, Azrai M, KasimF, and Mejaya MJ, 2007. Prospects for using molecular markers in corn breeding programs. Maros (ID): Food Crop Research and Development Center, Balitsereal.
- Petr FC and Frey KJ, 1966. Genotypic correlation, dominance, and heritability of quantitative characters in oats, *Crop Sci.* 6:259-262.
- Poespodarsono S, 1988. Plant Breeding Basics, Cooperate with IPB Information Resource Agencies, Bogor, Indonesia. pp :163.
- Stansfield, WD, 1991. *Genetica* second edition, Erlangga, Jakarta, Indonesia. pp :112-115.
- Subandi MS and Pasaribu D, 1996. Report on Results of Monitoring of Diseases and Seeds in Hybrid Maize Planting, Internal Report of the Food Crop Research and Development Center, Bogor, Indonesia.
- Suryo, 2008. *Genetica : Strata I*. Yogyakarta: Gadjah Mada University Press. Indonesia. Pp : 320.
- Takdir AM, Iriany RNM, Dahlan MM, Baihaki A, Rostini N, and Subandi, 2003. Genetic Control of Maize Resistance to Downy Mildew, *Food Crop Agricultural Research*, 22 (2) : 101-105.
- Talanca AH, 2010. Status of Vesicular Arbuscular Mycorrhizal Fungi (MVA) in Plants. Proceedings of the National Cereal Week, 353-357.
- Talanca AH, 2009. Resistance of Varieties / Corn Germplasm for Downy mildew. Proceedings of the National Seminar and Workshop, Sustainable Agricultural Technology Innovation Supporting Rural Agribusiness and Agro-Industry Development. Ministry of Agriculture, Agency for Agricultural Research and Development, Center for Assessment and Development of Agricultural Technology, Bogor. pp : 21-26.
- Te-Chato S, Lim M and Masahiro M, 2005. Comparison of Cultivar Identification Methods of Longkong, Langsat and Duku : *Lansium spp.* *Songklanarin J. Sci. Technol.*, 27 (3) : 465-472.
- Yasin HG, Rahman MA, Subekti NA, 2008. The General Combining Ability and Specific Combining Ability of lines of high protein maize expectations, *Journal of Food Crops Research Journal* 27(2) : 76-80.