

Available online freely at www.isisn.org

Bioscience Research

Print ISSN: 1811-9506 Online ISSN: 2218-3973 Journal by Innovative Scientific Information & Services Network



RESEARCH ARTICLE BIOSCIENCE RESEARCH, 2020 17(1): 347-350.

OPEN ACCESS

Chitosan edible coating and vacuum dehydration for preservation of dried *Artemia sp.* Cyst

Minh Phuoc Nguyen

Faculty of Biotechnology, Ho Chi Minh City Open University, Ho Chi Minh City, Vietnam

*Correspondence: minh.np@ou.edu.vn Received 12-01-2020, Revised: 22-02-2020, Accepted: 24-02-2020 e-Published: 01-03-2020

Fresh *Artemia* cysts have been considered a good source of proteins, lipids and carbohydrate. However their soft body are highly perishable during post-harvest owing to high water content, proteolytic enzyme. Therefore, appropriate preservation methods are necessary to maintain a good quality of this biomass. Chitosan has been well known with a broad range of applications in the food industry. Vacuum drying is one of the most energy demanding processes. Water evaporation also takes place at lower temperatures under vacuum, and hence the product processing temperature can be significantly lower, offering higher product quality. This study investigated on the possibility of chitosan coating in different concentration (0.5%, 1.0%, 1.5%, 2.0%, 2.5%) and vacuum drying under various pressures (-0.2, -0.4, -0.6, -0.8, -1.0 bar). Our result revealed that 2.0% chitosan coating and -0.8 bar vacuum drying were adequate to maitain the most *eicosa pentaenoic acid* (EPA) content during dehydration of artemia biomass. From this approach, the best valuable component inside the dried artemia cyst would be preserved effectively.

Keywords: Artemia, cyst, chitosan, coating, vacuum, drying, eicosa pentaenoic acid

INTRODUCTION

Artemia spp. are known as brine shrimp which are typical inhabitants of extreme saline biotopes (Camargo et al., 2005). Artemia produces cysts that float on the water surface (Hossein Tajik et al., 2008). These cysts are rich of protein, carbohydrate, total lipids, ash, linolenic acid and eicosapentaenoic acid potential use in aquaculture as a suitable live food for postlarval shrimp and fish (Hachem Ben Naceur et al., 2012; Naegel and Rodriguez-Astudillo, 2004: Anh et al., 2009; Peykaran Mana N. et al., 2014). Artemia cysts contains a high moisture content with proteolytic enzymes so it's essential to preserve this biomass by appropriate preservation methods (Sorgeloos et al., 2001). The dried Artemia cysts could be stored for long shelf-life with various benefits over to fresh or frozen forms (Chua and Chou 2005; Kamalakar et al., 2013; Anh et al.,

2014). The level of essential fatty acids such as eicosa pentaenoic acid (EPA) and docosahexaenoic acid (DHA) are a very important factor to determine the dietary value of *Artemia* cysts.

Chitosan is a de-acetylated form of chitin obtained from the crustacean waste (Santosh Kumar et al., 2019). Chitosan has attracted attention for food applications owing to its superior characteristic properties, such as degradability, solubility in weak acids. pH-sensitivity, filmbiodegradability. formina property, non-antigenic, biocompatibility, absence of toxicity, and low-cost (Bonilla et al., 2014; Croisier et al., 2013; Kumari et al., 2014; Santosh Kumar et al., 2019). Vacuum drying is more beneficial than hot air drying, because of short drying time, high drying rate and superior quality of dried products (Liliana Seremet et al., 2016). Vacuum

drying is an important process for drying highly heat-sensitive materials. The use of vacuum drying lowers the solvent boiling temperature, permitting operation at lower temperatures, directly influencing final product quality (Péré and Rodier, 2002). Vacuum-dried materials are characterized by better quality retention of nutrients and volatile aroma (S. K. Giri et al., 2014). Pupose of our study penetrated on the chitosan coating as pretreatment and vacuum drying to EPA content of the dried artemia cysts.

MATERIALS AND METHODS

2.1 Material

Fresh Artemia biomass (cysts) was collected from Bac Lieu province, Vietnam. After collecting, they must be kept in ice chest and quickly transferred to laboratory for experiments. Chemical substances and reagents such as chitosan powder, acetic acid were all analytical grade supplied from Rainbow Trading Co. Ltd., Vietnam.

2.2 Researching procedure

Experiment #1: Fresh Artemia cyst was encapsulated in chitosan coating in different concentrations (0.5%, 1.0%, 1.5%, 2.0%, 2.5%) and then dried by vaccum drier at 40°C under ambient pressure for 4 hours. The dried Artemia cyst was then analyzed the eicosa pentaenoic acid (EPA) and docosahexaenoic acid (DHA) content to determine the right edible coating.

Experiment #2: Fresh Artemia cyst was encapsulated in 2.0% chitosan coating and then dried by vaccum drier at 40°C under different pressure (-0.2, -0.4, -0.6, -0.8, -1.0 bar) for 4 hours. The dried Artemia cyst was then analyzed the eicosa pentaenoic acid (EPA) and docosahexaenoic acid (DHA) content to determine the right vacuum drying pressure.

2.3 Antioxidant capacity and statistical analysis

Eicosa pentaenoic acid (EPA, mg/g) and docosahexaenoic acid (DHA, mg/g) were **Table 1: Effect of chitosan concentration in** dehydration determined by high-performance liquid chromatography with electrochemical detection (Kotani A et al., 2016). The experiments were run in triplicate with three different lots of samples. Statistical analysis was performed by the Statgraphics Centurion XVI.

RESULTS AND DISCUSSION

3.1 Effect of chitosan concentration in coating to fatty acids in *artemia* cyst during dehydration

The price of Artemia cysts was dependent on nutritional value especially the presence of the acid (EPA) eicosa pentaenoic and docosahexaenoic acid (DHA) content. These contents were significantly affected by the chitosan coating (shown in table 1). Among these treatments, we realized that 2.0% of chitosan was appropriate for preservation of these valuable fatty acids containing in the dried artemia cysts during dehydration. In relevent studies, a fluidized bed dryer for dehydration of Artemia cysts at 40°C was evaluated (T. Bosteels et al., 1996). One study evaluated the effect of different drying techniques at different temperatures on the contents of total lipid and fatty acid profile of Artemia biomass. The intermittent microwave combined with convective hot air drying was a promising technique, which could produce high quality dried products in short drying times (Nguyen Thi Ngoc Anh et al., 2015).

3.2 Effect of vacuum drying pressure to fatty acids in *artemia* cyst during dehydration

The DHA level, especially DHA/EPA has an importan role in growth, survival rate and protection against diseases in marine fish and crustacean (Watanabe, 1993; Furuita, 1996; Han et al., 2001). In our research, fatty acid profiles showed a significant difference by vacuum drying pressure (shown in table 2). It's obviously noticed that -0.8 bar was ideal for dehydration of artemia cyst under vacuum.

Table 1: Effect of chitosan concentration in coating to fatty acids in *artemia* cyst during dehydration

Parameter	Chitosan concentration (%)								
	Control	0.5	1.0	1.5	2.0	2.5			
EPA (mg/g)	14.25±0.03°	19.78±0.01 ^b	20.37±0.01 ^{ab}	21.49±0.01 ^{ab}	22.63±0.00 ^a	22.70±0.02 ^a			
DHA (mg/g)	0.37±0.00 ^c	0.65±0.02 ^b	0.78±0.03 ^{ab}	0.89±0.00 ^{ab}	0.95±0.03 ^a	0.96±0.01 ^a			

Note: the values were expressed as the mean of three repetitions; the same characters (denoted above), the difference between them was not significant ($\alpha = 5\%$).

Parameter		Vacuum drying pressure (bar)						
	Control	-0.2	-0.4	-0.6	-0.8	-1.0		
EPA (mg/g)	22.63±0.00°	22.79±0.03bc	22.84±0.02 ^b	22.96±0.03 ^{ab}	23.05±0.02 ^a	23.06±0.00 ^a		
DHA (mg/g)	0.95±0.03°	1.03±0.01 ^{bc}	1.09±0.03 ^b	1.14±0.01 ^{ab}	1.19±0.00 ^a	1.20±0.00 ^a		

Table 2: Effect of vacuum drying pressure to fatty acids in artemia cyst during dehydration

Note: the values were expressed as the mean of three repetitions; the same characters (denoted above), the difference between them was not significant ($\alpha = 5\%$).

CONCLUSION

The content of highly unsaturated fatty acids like EPA and DHA is a vital parameter that defines the nutritional composition and commercialization of *Artemia* syst for shrimp, marine larvae, fish aquaculture. These fatty acids are highly affected by different variables during processing. We have successfully investigated some major factors influencing to the stability of EPA and DHA content in *artemia* syst throughout the dehydration. In the dried form, shelf-life of artemia syst can be extended for a long period of time.

CONFLICT OF INTEREST

The authors declared that present study was performed in absence of any conflict of interest.

AUTHOR CONTRIBUTIONS

This is a single author publication.

Copyrights: © 2020@ author (s).

This is an open access article distributed under the terms of the **Creative Commons Attribution License (CC BY 4.0)**, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author(s) and source are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

REFERENCES

- Anh, N.T.N., Hien, T.T.T., Wille, M., Hoa, N.V. and Sorgeloos, P. (2009). Effect of fishmeal replacement with *Artemia* biomass as protein source in practical diets for the giant freshwater prawn *Macrobrachium rosenbergii. Aquaculture Research* 40: 669-680.
- Anh, N.T., Nhi, N.T. and Hoa, N.V. 2014. Comparative study of solar and sun drying of *Artemia* biomass in the Mekong delta of Vietnam. *Journal of Science and Technology*

52: 336-342.

- Bonilla, J.; Fortunati, E.; Atarés, L.; Chiralt, A.; Kenny, J.M. (2014). Physical, structural and antimicrobial properties of poly vinyl alcohol– chitosan biodegradable films. *Food Hydrocoll.* 35: 463–470.
- T. Bosteels, W. Tackaert, G. Van Stappen and P. Sorgeloos (1996). Improved use of the fluidized bed dryer for Artemia cysts. *Aquacultural Engineering* 15: 169-179.
- Camargo, W.N.; Duran, G.C.; Rada, O.C.; Hernandez, L.C.; Linero, J.C.; Muelle, I.M.; Sorgeloos, P. (2005). Determination of biological and physicochemical parameters of *Artemia franciscana* strains in hypersaline environments for aquaculture in the Colombian Caribbean. *Saline Systems* 26: 1-9.
- Chua, K.J. and Chou, S.K. (2005). A comparative study between intermittent microwave and infrared drying of bioproducts. *International Journal of Food Science and Technology* 40: 23-29.
- Croisier, F.; Jérôme, C. (2013). Chitosan-based biomaterials for tissue engineering. *Eur. Polym. J.* 49: 780–792.
- Furuita, H., Takeuchi, T., Toyota, M. and Watanabe, T., 1996. EPA and DHA requirements in early juvenile Red Sea bream using HUFA enriched Artemia nauplii. *Fisheries Science* 62: 246–251.
- S. K. Giri, P.P. Sutar and Suresh Prasad (2014). Effect of process variables on energy efficiency in microwave-vacuum drying of button mushroom. *Journal of Food Research and Technology* 2: 31-38.
- Hachem Ben Naceur, Amel Ben Rejeb Jenhani and Mohamed Salah Romdhane (2012). Quality characterization of cysts of the brine shrimp *Artemia salina* from Tunisia focusing on their potential use in aquaculture. *Journal of Biological Research-Thessaloniki* 17: 16 – 25.
- Han, K., Geurden, I. and Sorgeloos, P., 2001. Fatty acid changes in enriched and subsequently starved *Artemia franciscana*

nauplii enriched with different essential fatty acids. *Aquaculture* 199: 93-105.

- Hossein Tajik, Mehran Moradi, Seyed Mehdi Razavi Rohani, Amir Mehdi Erfani and Farnood Shokouhi Sabet Jalali (2008). Preparation of chitosan from brine shrimp (*Artemia urmiana*) cyst shells and effects of different chemical processing sequences on the physicochemical and functional properties of the product. *Molecules* 13: 1263-1274.
- Kamalakar, D., Nageswara, L.R., Rao, K.S.P. and Venkateswara, M.R. (2013). Studies on drying characteristics of prawn and fish. *Journal of Chemical, Biological and Physical Sciences* 3: 972-982.
- Kumari, S.; Rath, P.K. (2014). Extraction and characterization of chitin and chitosan from (*Labeo rohit*) fish scales. *Procedia Mater. Sci.* 6: 482–489.
- Naegel, L.C.A. and Rodriguez-Astudillo, S. (2004). Comparison of growth and survival of white shrimp postlarvae (*Litopenaeus vannamei*) fed dried Artemia biomass versus four commercial feeds and three crustacean meals. *Aquaculture International* 12: 573-581.
- Nguyen Thi Ngoc Anh, Nguyen Thuan Nhi and Nguyen Van Hoa (2015). Effect of different drying methods on total lipid and fatty acid profiles of dried *Artemia franciscana* biomass. *Can Tho University Journal of Science* 1: 1-9.
- Péré, C., Rodier, E. (2002). Microwave vacuum drying of porous media: experimental study and qualitative considerations of internal transfers. *Chemical Engineering and Processing* 41: 427-441.
- Peykaran Mana N.; Vahabzadeh H.; Seidgar M.; Hafezieh M. Pourali H. R. (2014). Proximate composition and fatty acids profiles of Artemia cysts, and nauplii from different geographical regions of Iran. *Iranian Journal of Fisheries Sciences* 13: 761 -775.
- Santosh Kumar, Fei Ye, Sergey Dobretsov and Joydeep Dutta (2019). Chitosan nanocomposite coatings for food, paints, and water treatment applications. *Appl. Sci.* 9: 2409.
- Sorgeloos, P., Dhert, P. and Candreva, P. (2001). Use of the brine shrimp, *Artemia* spp., in marine fish larviculture. *Aquaculture* 200: 147-159.
- Kotani A, Watanabe M, Yamamoto K, Kusu F, Hakamata H. (2016). Determination of

eicosapentaenoic, docosahexaenoic, and arachidonic acids in human plasma by highperformance liquid chromatography with electrochemical detection. *Anal Sci.* 32:1011-1014.

- Liliana Seremet, Elisabeta Botez, Oana-Viorela Nistor, Doina Georgeta Andronoiu, Gabriel-Danut Mocanu (2016). Effect of different drying methods on moisture ratio and rehydration of pumpkin slices. *Food Chemistry* 195: 104-109.
- Watanabe, T., 1993. Importance of docosahexaenoic acid in marine larval fish. *Journal of World Aquaculture Society* 24: 152–161.